

Latest advances in combining gold nanomaterials with physical stimuli towards new responsive therapeutic and diagnostic strategies

¹Dania Movia*, ^{2,3}Maroua Benhaddada, ²Jolanda Spadavecchia, ^{1,4}Adrielle Prina-Mello*
 1 LBCAM, Department of Clinical Medicine, Trinity Translational Medicine Institute, Trinity College Dublin, Dublin, Ireland

2 CNRS, UMR 7244, NBD-CSPBAT, Laboratoire de Chimie, Structures et Propriétés de Biomatériaux et d'Agents Thérapeutiques Université Sorbonne Paris Nord, Bobigny, France

3 TORSKAL nanoscience, Sainte Clotilde, La Réunion, France

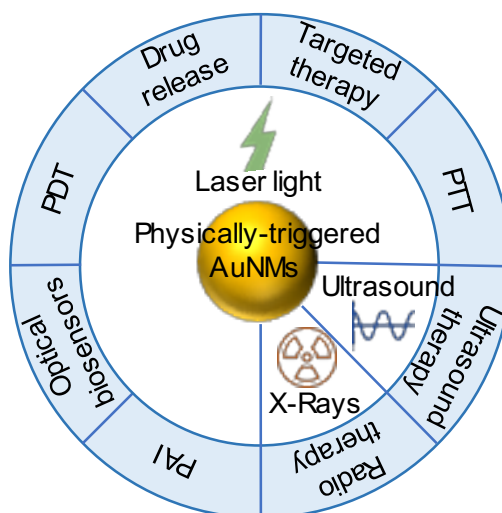
4 AMBER Centre, CRANN Institute, Trinity College Dublin, Dublin, Ireland

Submitted: March 18, 2020

Accepted: April 19, 2020

Published: April 22, 2020

Graphical abstract



Keywords

Gold nanoparticle¹; external stimuli; physical stimuli; physically triggered nanomedicine.

Abstract

Nanomedicine aims at enhancing treatment efficiency and/or improving diagnostic sensitivity by better controlling several critical parameters, such as tissue targeting and off-target toxicity. More recently, advanced nanomedicine products have been developed to achieve spatially and temporally controlled therapy and diagnosis. This review focuses on gold nanomaterials (AuNMs) and alloy/hybrid AuNMs that can be used in stimuli-responsive strategies for therapeutic and diagnostic applications. Endogenous and/or exogenous stimuli can be used as a trigger for such systems. Herein, we focus on those activated by exogenous stimuli. Our review starts from one specific externally activated product, Aurolase®, which recently underwent clinical studies. Further we continue describing a specific physically triggered category, for which the exogeneous stimulus applied induces a structural transformation or modification that is essential for their therapeutic and/or diagnostic action. Gold nanomaterials are grouped both by the nature of the function they exert (therapeutic or diagnostic) and the stimuli class.

* Corresponding authors: Email: dmovia@tcd.ie; prinamea@tcd.ie; Full postal address: Lab 0.74, Trinity Translational Medicine Institute, Trinity Centre for Health Sciences, James's Street, D8, Dublin, Ireland.

¹ In this review we will refer to nanomaterials without detailing their geometrical shapes since those of secondary importance to the scope of the review.

Rationale and Purpose

In the last decades, the growing incidence of chronic diseases associated with the increased aging of the population has triggered the need for more effective and localized therapies as well as efficient diagnostic tools for early disease detection. AuNMs and alloy/hybrid AuNMs have been widely investigated to address this need. The gold nanomaterials reviewed herein constitute the most advanced research on this promising technology.

This review focuses on those AuNMs and alloy/hybrid AuNMs activated by externally applied physical stimuli.

Introduction

Clinical translation of AuNMs to date

Gold nanomaterials (AuNMs) and alloy/hybrid AuNMs offer a valuable platform for a wide range of biomedical applications, ranging from therapy to diagnosis [1, 2].

Furthermore, both AuNMs and alloy/hybrid AuNMs provide excellent platforms to develop multifunctional approaches (“theragnostics”) [3-5]. As compared to gold nanomaterials, alloy or hybrid AuNMs offer the added advantage of combining the unique properties of multiple components within a single product. Alloy AuNMs incorporate inorganic components (e.g., iron oxide [6, 7]); whereas, hybrid AuNMs integrate a gold core within organic nanostructures (polymeric nanoparticles [8], liposomes, dendrimers, micelles or biomacromolecules [9]). Owing to their composition, alloy and hybrid AuNMs provide excellent platforms to develop theragnostic tools, where the nanomaterial is used as diagnosis probe and therapeutic agent at the same time.

Various gold and alloy/hybrid gold nanoparticles are currently under investigation in clinical studies for their application as therapeutic and/or diagnostic tools (Table 1). However, no AuNM has reached clinical approval for therapeutic applications [10-12]. Conversely, examples of FDA-approved, AuNM-based diagnostics can be found, such as the Uni-Gold Recombigen HIV-1/2, a single-use rapid immunoassay for the qualitative detection of antibodies to human

immunodeficiency virus (HIV)-1 and/or HIV-2 in human serum, plasma, and whole blood [13]; the VERIGENE® system, for the detection of DNA, RNA or protein targets; the First Response Gold Digital Pregnancy Test [14]; and the Single Path *E. Coli* O157 Gold Labelled Immunosorbent Assay, used in food analysis laboratories. All these diagnostic tests fall under the optical biosensor category, where AuNMs are used as labels for signal amplification in bio-recognizing events

AuNMs and alloy/hybrid AuNMs can be finely tuned so that their therapeutic effect or diagnostic action is triggered by specific stimuli [15-17]. Stimuli can be associated with specific biological (endogenous) factors, such as the microenvironmental properties of a defined diseased condition (e.g., acidic pH [18] and levels of reductive agents [19] in the tumour tissue, or intracellular enzymatic activity [20]). These are the so-called “internal stimuli” or “intrinsic stimuli” and are generally associated with disease progression. Alternatively, AuNMs can be designed to respond to externally applied physical stimuli (“external stimuli” or “exogeneous stimuli”): examples include, but are not limited to, light, temperature, magnetic and electric field, and X-rays. As compared to internal triggers, external stimuli are easier to control, are not associated with a patient- or disease-dependant variability and can be selected based on key parameters like cost and safety.

Aurolase® Therapy – An example of physical stimuli-triggered AuNMs from the clinics

Among the AuNMs under clinical development listed in Table 1, the Aurolase® therapy is of particular interest for this review, as it is a unique example of a stimuli responsive AuNM that has been translated from the lab bench to the clinic. No other stimuli-responsive, AuNM-based therapy has entered clinical trials yet. Similarly, no clinical example can be found of on/off AuNM switch probes that exert their detection function by responding to a physical stimulus. Diagnostic AuNM-based tools that have reached regulatory approval or are in a clinical trial to date act as signals enhancers.

Table 1. Ongoing clinical studies on gold and gold alloy/hybrid nanomaterials found on clinicaltrials.gov (last updated: February 2020).

Application	Product name	Product type	Condition or disease	Clinical study phase	ClinicalTrials.gov Identifier	
Therapy	Drug delivery	NU-0129	SNA bound on spherical AuNPs	Gliosarcoma Recurrent glioblastoma	Early Phase 1	NCT03020017
		C19-A3 GNP (also known as MTX 102)	Intradermal micro-injectable solution of human C19A3 proinsulin peptide coupled to carbohydrate-coated gold nanoparticles	Type 1 diabetes	Phase 1	NCT02837094
	Induce re-myelination	CNM-Au8	Gold nanocrystals	Chronic optic neuropathy in multiple sclerosis	Phase 2	NCT03536559
				Parkinson's disease	Phase 2	NCT03815916
	Photo-thermal ablation therapy	AuroLase® Therapy	Auroshells® (silica nanoparticles with a gold shell)	Prostate neoplasm	N/A	NCT02680535
						NCT04240639
Cavity surface pre-treatment	Nano Care Gold	Mixture of AuNPs and AgNPs suspended in 70 % isopropyl alcohol	Caries	N/A	NCT03669224	
Diagnosis	Contrast agent	SEB-250	Silica nanoparticles surrounded by a shell of gold	Acne vulgaris	N/A	NCT03573115
	Artificial nose	AuNPs	Cross-reactive AuNPs coated with organic ligands	Pulmonary arterial hypertension	N/A	NCT02782026

The clinical study phase is indicated as “not applicable” when referring to studies without FDA-defined phases, such as trials of medical devices. Only active clinical trials are listed; completed or terminated clinical trials are not reported.

Abbreviations: AgNPs = silver nanoparticles; AuNPs = gold nanoparticles; SNA = spherical nucleic acid.

The AuroLase® product, developed by Nanospectra, is based on the use of silica-gold nanoshells, called Auroshells™, coated with (poly)ethylene glycol (PEG) and with sizes between 140 and 150 nm. Auroshells™ are designed to thermally ablate the solid tumors

following stimulation with a NIR high-power diode source as external stimuli [21-23].

At the pre-clinical stage, Auroshells™-mediated photothermal therapy has been reported to improve the overall survival in various cancer models, including for example a murine xenograft model of glioma [24] and an

orthotopic canine model of a brain tumor [25]. The safety profile for the intravenous administration of Auroshells™ was proven in vitro [26], in vivo, [26] and patients with human prostate cancer [27]. Once intravenously administered, progressive extravasation and accumulation of Auroshells™ was demonstrated at the tumor site [28], associated with the enhanced permeation and retention (EPR) effect [28, 29]. In this context, human data showed that the clinical use of Aurolase® therapy has one main limitation: the EPR-driven accumulation of the Auroshells™ is heterogeneous within tumor tissues. The EPR effect is a passive targeting mechanism [29] that refers to the preferential accumulation and site-specific delivery of nanomaterials in the perivascular tumor region, due to the leakiness of the tumor vasculature and poor lymphatic drainage [30]. Briefly, systemically administered nanomaterials cross the tumor vascular barrier through intercellular gaps, are retained within the tumor tissue owing to the pressure created by poor lymphatic drainage, and penetrate by convection [31] through the tumor stroma to reach the cancer cells [32]. However, the tumor vasculature is characterized by abnormal angiogenesis, which leads a heterogeneous vessel network formation and, subsequently, tumor areas where the vasculature is completely lacking [33]. This, along with the high interstitial pressure and the mechanical stresses generated by the interactions between cancer/stromal cells and the extracellular matrix, lead to a heterogeneous EPR effect that negatively affects nanoparticle accumulation in the tumor [33]. Furthermore, large (> 50 nm in size) nanoparticles such as Auroshells™ have been shown to poorly penetrate dense tumor stroma [33]. The clinical consequence is that the highest Auroshells™ doses are achieved in the tumor cortex, while almost no accumulation is detectable within the necrotic tumor core [28].

In a first clinical trial (NCT00848042), the Aurolase® therapy was used in the treatment of patients with refractory and/or recurrent tumors of the head and neck. Three treatment groups of five patients each were enrolled and observed for six months following treatment. Each group received a single dose of AuroShells™ followed by one or more interstitial illuminations with a NIR laser. Particle dose and laser power differed among

groups: the AuroShells™ doses tested were equal to 4.5 and 7.5 mL/Kg; whereas, laser power ranged between 3.5 to 5 watts. The therapy was shown not to be particularly safe, with serious and other side effects developed by many patients and attributable to the administration of the nanoparticles (NCT00848042). A second trial (NCT01679470) tested the efficacy of Aurolase® therapy for the localized treatment of primary or metastatic lung tumors with airway obstruction. NIR irradiation was delivered by optical fibre via bronchoscopy. This trial was, however, terminated. The most recent trial (NCT04240639), posted in January 2020 and currently recruiting patients, uses AuroShells™ as a contrast agent for magnetic resonance imaging (MRI), coupled to real-time transrectal ultrasound (US) to direct nanoparticles-triggered focal ablation of prostate neoplasms using NIR laser irradiation generated by means an FDA-approved laser and an interstitial optical fiber diffuser. This trial was approved based on very promising in vitro [34] and in vivo [35] data. According to the trial design, up to sixty prostate cancer patients will be recruited and will receive a single intravenous infusion of AuroShells™ 12 to 36 hours before MRI/US-guided laser irradiation. Tumor ablation will be assessed: (i) by contrast-enhanced MRI at 48 to 96 hours after laser illumination; (ii) by MRI/US-guided target biopsy at 6 months after laser treatment; and (iii) MRI/US-guided target biopsy in combination with standard biopsy after 1 year. NCT04240639 trial is expected to be completed by June 2023.

Discussion

As mentioned above, Aurolase® therapy is the sole stimuli responsive AuNM that has been translated from the lab bench to the clinic. Nevertheless, the scientific literature includes many other examples of AuNMs that respond to stimuli. Recently, a review by Tian et al. [16] reported on the main externally-stimulated AuNMs products that are at the early-stage development phases. Our review focuses instead on the means of external physical stimuli that can be employed in combination with AuNMs, and how these stimuli can induce modifications in the nanomaterial structure to trigger a therapeutic or diagnostic action, as the most advanced research on this promising

technology. Modifications in the nanomaterial structure include both chemical and physical changes, such as, for example, light-induced cleavage of photosensitive chemical bonds for spatial- and temporal-controlled drug release, melting of thermoresponsive coating polymers and subsequent targeted drug release, and nanomaterial phase transitions or polymer degradation in response to ultrasound stimulus. For more details on the design strategies that can be used for programming physical stimuli-responsive nanomaterials, a comprehensive review has been recently published by Sahle et al. [36].

The following sections discuss the types of external stimuli that can cause structural modifications in AuNMs, and how these chemical or physical modifications can be used for both therapeutic and diagnostic applications.

Types of triggers for stimuli responsive AuNMs

Non-ionizing radiations as external physical stimuli

Laser light

Laser light is the most common non-ionizing radiation used as external stimuli for AuNMs and alloy/hybrid AuNMs. Light is characterized by clinical relevance, excellent spatio-temporal controllability and indeed clinical safety. Its use is associated with AuNMs ability to efficiently absorb light, a property that results from the localized surface plasmon resonance (LSPR) effect. When administering light at a frequency that overlaps with the LSPR absorption, this is efficiently scattered generating both linear and nonlinear (e.g., third-harmonic generation) optical processes [37]. The remaining part of the energy is absorbed and dissipated into heat, through a rapid photothermal conversion by electron-phonon interaction [38, 39]. Photothermal conversion can lead not only to temperature increase but also to the generation of US waves and acoustic cavitation [39].

Targeting and functionalization strategies do not affect the photo-induced behaviour of AuNMs, enabling the multimodal/theragnostic applications of physical stimuli-triggered gold nanomaterials. On the other hand, the LSPR wavelength is influenced by several AuNM properties, including nanomaterial size, shape,

structure, composition and the dielectric constant of the surrounding medium [40]. Increases in AuNMs size and aspect ratio, for example, result in a plasmonic resonance shift from an ultraviolet (UV)-visible range (400–520 nm) to the near-infrared (NIR) region (800–1200 nm) [16, 39]. For maximizing the effect of the physical stimulus, the laser light wavelength should be selected so that to match that of the LSPR band. Thus, the possibility to tune the LSPR of AuNMs towards the NIR light window is extremely advantageous, as tissues and biological fluids are moderately transparent in such spectral region, commonly known as “water window.” In theory, UV light can also be employed as a physical stimulus, but it is associated with a limited ability to reach deep-seated tissues within the body. In general, small spherical AuNMs have higher photothermal conversion efficiency than those with larger diameters [41]. Furthermore, non-spherical AuNMs, including nanorods and nanostars, are more efficient than spherical AuNMs in producing heat following light stimuli, due to their larger absorption cross sections [42].

Temperature

Changes in temperature are used as an external stimulus to induce phase transitions in a controlled manner. Temperature-responsive systems are mainly hybrid AuNMs, where the gold core is coupled to a thermoresponsive polymer, such as poly(N-substituted acrylamide)s or poly(N-isopropylacrylamide-co-2-(dimethylamino)-ethylmethacrylate) [43]. Coupling is obtained via copolymerization, conjugation and/or grafting [36]. The intensity of the temperature stimulus needed to trigger a response, therefore, depends on the polymer type used and is independent of the AuNM physico-chemical properties.

Electric pulse and electric field

Electric waves are another external stimulus that can be used to activate the AuNMs therapeutic action. Electrical stimuli are relatively easy to generate, control, and remotely apply without the need for sophisticated instruments, which makes electro-responsive AuNMs a very attractive system.

Electric waves can be used to generate electroporation, where electrical pulses have been employed, for example, to achieve gene release from electro-responsive AuNMs in

animal models [44]. Electric waves can also generate an electric field, which triggers local heating of AuNMs [45, 46]. Because of differences in the characteristics of the AuNMs tested so far, the research community has not reached a consensus on the mechanism determining AuNM heat generation using an electric field. Nevertheless, two parameters have been identified as critical: electric field source and AuNMs properties, including size, concentration, and aggregation [47].

Ultrasound (US)

US waves are sound waves with frequencies higher than the upper audible limit of human hearing. US has been utilized as an exogenous stimulus for biomedical applications due to its non-invasiveness, ease of accessibility, cost-effectiveness, lack of ionizing radiation residues, controllable spatio-temporal effect, and high patient acceptability.

AuNMs can be activated by US waves in two ways [48]. They can be used together with sensitizing agents [49, 50], to prolong the non-radiative relaxation time of the sensitizer, thus promoting the generation of singlet oxygen. AuNMs can also act as nanosensitizers themselves [51, 52]. In the latter case, AuNMs act by enhancing the inertial cavitation rate in the biological tissue [53-55].

Magnetic field

Magnetic fields have long been investigated as an external stimulus for magnetic nanoparticles. However, AuNMs are not magnetic. In 2008, the concept of “super atom” has been used for the first time to indicate the extraordinary ferromagnetic and paramagnetic properties of small gold clusters [56]. These properties were then further described by other groups in the following years [57-59]. A current hypothesis explaining the mechanism by which AuNMs show magnetic properties is still controversial and has been extensively discussed elsewhere [60, 61]. However, applications of gold clusters for therapeutic hyperthermia under alternative magnetic force exposure have not been developed yet [62]. On the other hand, alloy iron oxide-gold nanoparticles have been investigated as contrast agents for MRI by exploiting the magnetic properties of iron oxide [63-67]. Magnetic alloy AuNMs and their biomedical applications have been comprehensively described in a recent review [68].

Ionizing radiations as external physical stimuli

X-rays

Gold is a high-Z element ($Z=79$) that has been widely used as radiosensitizers owing to its good absorption of ionizing radiations. Radiosensitization induces reactive oxygen species (ROS) production, increases oxidative stress, and promotes DNA damage in the targeted tissue by chemical interactions.

Like gold, AuNMs can be used to improve radiosensitivity [69]. The most famous example of AuNMs-based radiosensitizer is AuroVist™, a commercially available product for in vivo and in vitro research. In AuNMs radiosensitizers, the incident X-ray wave interacts with the nanoparticles, resulting in the emission of secondary electrons. Such electrons cause cell damage by direct interaction (“physical enhancement”), production of free radicals (“chemical enhancement”), and/or production of ROS and oxidative stress (“biological enhancement”). Detailed reviews describing the radiosensitization mechanisms triggered by AuNMs have recently published [70-72]; whereas, Cole et al. have discussed the nanomaterial optimized properties, such as composition, mass concentration, size, shape and surface functionalization, that ensure AuNM optimal functional performance as contrast agents in X-ray imaging and computed tomography [73].

In addition to radiosensitization, X-rays can be used as physical stimulus in conjunction with AuNMs to trigger specific therapeutic actions (e.g., drug release) or to allow for medical imaging by, for example, computed tomography (CT).

Stimuli-responsive AuNMs modifying their structure following an external physical trigger

Therapeutic applications

A schematic summary of the AuNMs described in this section is presented in Table 2.

Photo-responsive drug release

Light sources have been used in pre-clinical research to release therapeutic payloads (e.g., drugs, genes) from AuNMs employed as carriers [74]. This strategy offers the advantage to allow for the controlled release of the therapeutic active compound at a specific location and in a controlled manner. The

therapeutic dosage needed is therefore reduced and the side effects diminished, positively impacting on the treatment outcomes and safety [75]. An extensive literature review has been recently published on this topic [76].

The main mechanism by which photo-triggered drug delivery is achieved is the induction of AuNMs heating via the LSPR effect [77]. The photo-induced particle heating can be used to cause the thermal disruption of the non-covalent interactions between the payload molecules and AuNMs. Such structural changes are triggered via photo-caging or photo-isomerization [78]. These approaches have been used to affect the hydrophilic-hydrophobic balance of thermally responsive compounds coating the AuNMs surface (e.g., polymers or DNA [79-81]) or entrapped within the AuNMs, thus triggering the release of the therapeutic payload. For example, the LSPR photothermal effect was successfully used to release a chemotherapeutic drug (paclitaxel) loaded within poly(lactic acid-co-glycolic acid) (PLGA) microspheres that contained hollow gold nanospheres [82].

NIR light has been employed to induce the release of doxorubicin from hollow AuNMs via the LSPR photothermal effect [83, 84]. Also, gold nanospheres of 3–7 nm incorporated into the lipid bilayer of liposomes, have been used to fabricate photothermally responsive hybrid AuNMs capable of releasing fluorescein in a controlled manner [85]. Ma et al. incorporated gold nanoshells, doxorubicin, and magnetic nanoparticles into micelles made from cholesteryl succinyl silane. Upon laser irradiation, the photothermal heating of the gold nanoshells induced the micelles to collapse and resulted in doxorubicin release [63]. This photothermal approach can be used to induce deformation and rupture of gold nanoshells, where the therapeutic payload has been encapsulated, thus triggering its controlled release [84, 86-88]. A mechanism describing how drugs are released from these AuNMs when combined with laser-induced thermal therapy has also been proposed [89]. An alloy photo-responsive system formed by mesoporous silica-coated gold nanorods, was reported to be able to release doxorubicin in a stimuli-controlled manner [90]. Finally, a photo-responsive drug delivery system for the treatment of age-related macular degeneration, an ocular disease, was also developed based on

polymer-coated AuNMs entrapped within an agarose hydrogel containing bevacizumab as therapeutic payload [91]. A localized temperature increase was induced by exposure to visible light, which caused hydrogel softening and subsequent drug release. These studies demonstrate that polymeric matrixes do not prevent light from reaching AuNMs and they do not compromise their photothermal properties. In parallel, photo-induced heating has been used to release oligonucleic acids from AuNMs via thermal dehybridization [92-96].

On the other hand, light irradiation can also induce payload release by mechanisms other than the LSPR photothermal effect, for example by disrupting the bonds that link the payload to the AuNMs surface. This drug delivery strategy is referred to as “light-triggered photocaged strategy,” and it exploits NIR-sensitive covalent bonds or UV-aided, photo-cleavable molecular gates to link the therapeutic molecule to the AuNMs. For example, Sreejivungsa et al. developed AuNMs featuring light-responsive linkers capable of releasing a model drug (a naturally occurring compound extracted from *Goniothalamus elegans Ast*) following exposure to UV light [97]. Once irradiated with UV light, the linker was cleaved, leading to the release of the therapeutic agent. Agasti et al. used a photo-responsive o-nitrobenzyl linkage that upon UV light irradiation dissociated, for the controlled release of a caged anti-cancer drug (5-fluorouracil, 5-FU) from AuNMs [98].

Photo-responsive targeted therapy

Light can be used not only to induce payload release but also to determine the timing and specific location at which AuNMs will bind to the targeted tissue, to improve efficacy and decrease systemic toxicity. For example, Yeh and co-workers also established a smart photo-activated AuNMs drug delivery system that, upon UV light activation, can selectively target tumor cells and release an anti-cancer drug (Taxol) [99]. NIR irradiation has also been used to recover the function of surface-bound ligands blocked by steric hindrance on alloy (silica-gold) nanoshells [100]. YIGSR peptides were bound to the surface of the AuNMs but were prevented from binding their target by a coating of thermoresponsive pNIPAAm-co-pAAm copolymer. Upon NIR irradiation, the gold nanoshells heated up, causing the collapse

of the copolymer and exposure of the peptide, allowing cell targeting.

Photothermal therapy (PTT)

PTT is defined as the photon-mediated increase of localized temperature that can stimulate physiological responses. In anti-cancer therapy, PTT is used to kill cancer cells via the generation of localized heat between 43°C and 49°C [101].

As previously mentioned, when irradiated with a NIR laser source, AuNMs can convert the light into heat, allowing their use as PTT agents [102-106]. Pitsillides et al. reported the first demonstration of the use of AuNMs as PTT contrast agents [107]. The mechanisms by which AuNMs generate heat upon light irradiation through non-radiative processes are described elsewhere [108, 109].

Temperature distribution within the tumor tissue is critical to the efficacy of PTT. AuNMs photothermal conversion is directly proportional not only to the incident laser power and irradiation duration but also to the size and shape of the nanomaterial [38, 110-112], as discussed in detail in a recent review published on this topic [113]. Another recent review summarizes the heat-transfer and thermal-damage models that can be used to estimate the influence of these parameters on heat distribution in the tumor tissue [114]. Among the various AuNMs types, gold nanorods have been reported to be the best contrast agent for PTT [42]. However, their penetration in the tumor tissue can be limited, negatively impacting the heat distribution throughout the tumor and, ultimately, the success of the PTT treatment. Spherical AuNMs better accumulate within the tumor tissue. However, tumor retention of small AuNMs is challenging because they are rapidly cleared from the extracellular milieu. Additionally, spherical AuNMs need to be more than 50 nm in size to effectively absorb in the NIR region. To overcome this size limitation, spherical AuNMs decorated with photolabile diazirine moieties, have been developed as PTT agents [115]. When exposed to UV irradiation, the diazirine moieties caused the AuNMs to aggregate in the tumor site, changing their size and resulting in a significant shift in their LSPR peak towards the NIR region, which in consequence induced

plasmonic coupling between adjacent AuNMs and remarkably enhanced their efficacy for PTT.

Photodynamic therapy (PDT)

PDT is another form of cancer treatment that utilizes light. Unlike PTT, which is oxygen-independent, PDT is completely dependent on the availability of oxygen in the tissue. In this technique, a photosensitizing agent (injected in the tissue) is excited at specific wavelengths, leading to the energy transfer, generation of ROS and cell death by apoptosis. AuNMs can act as photosensitizer agents. For example, Vijayaraghavan et al. developed lipid-coated gold nanoechinus (i.e., nanospheres with many spiky nanorods sticking out from the surface) that were capable of sensitizing tissues via singlet oxygen production for PDT applications [116].

Nevertheless, AuNMs have mainly been used as carriers to deliver photosensitizers to the target tissue [117, 118], thus overcoming the insolubility issue associated with the use of these compounds in physiological fluids. Gold nanostars have been used, for example, to deliver chlorin e-6, a PDT agent, allowing them to perform dual PDT/PTT therapy [119]. Other PDT/PTT dual therapies were developed by incorporating the photosensitizer hypocrellin into lipid vesicles containing gold nanocages [120], linking chlorin e6 to gold nanorods [121], or conjugating the photosensitizer indocyanine green onto gold nanospheres [122, 123]. Among stimuli-responsive AuNMs modifying their structure following light irradiation with application of PDT agents, we find gold nanorods conjugated to the photosensitizer AIPcS4 via electrostatic immobilization [124]. In this system, photothermal heating causes the release of the bound photosensitizer from the gold nanorod surface, allowing it to exert its PDT function.

High-intensity focused ultrasound therapy (HIFU)

When ultrasound is applied, the target tissue absorbs acoustic energy, triggering both thermal and non-thermal effects. In the thermal effects, tissue damage occurs as a function of the thermal increase. This is generally referred to as “sonodynamic therapy.” Because of the thermal effects, the cell membrane permeability

can also change, triggering endocytosis and enhancing the accumulation of the therapeutic within the cell. This process is called “sonoporation.” The use of AuNMs in sonoporation has been explored in pre-clinical research for enhanced drug delivery applications [125]. On the other hand, non-thermal effects include cavitation, such as the formation of microbubbles within the tissue; these interact with the ultrasound field, which induced them to oscillate, grow and eventually implode (a process called “inertial cavitation” or “transient cavitation”). The microbubble collapse is associated with a shock wave, shear stress, high temperatures and ROS formation that can mechanically and chemically damage the cancer tissue. The acoustic fluence required to produce cavitation is very high. HIFU [126-128], a therapeutic technique that uses a US intensity that is several orders of magnitude greater than that of standard US, is used to achieve the required acoustic fluence. However, internal cavitation is difficult to control. To overcome such limitation, researchers have been using AuNMs as agents to lower the threshold of cavitation intensity, acting both as cavitation nuclei and promoting the collapse of the microbubbles [49, 51, 129-135]. Also, AuNMs aggregation, which induces thermal field overlap and plasmonic coupling, has been investigated as a method to further lower the required fluence levels [136].

Focusing on physically triggered AuNMs or hybrid/alloy AuNMs whose structure is modified by the stimulus, PEGylated mesoporous silica nanocapsules, loaded with pyrene and perfluorohexane and functionalized on the surface with AuNMs, were formed [137]. In this alloy system, the AuNMs enabled for HIFU enhancement, which ultimately lead to the nanocapsule disruption and drug delivery, as demonstrated both in vitro and in vivo. Similarly, successful payload release was achieved from gold nanocages coated with the thermally responsive poly(NIPAAm-co-AAm) copolymer by exposure to HIFU [138]. In another study by Moon et al., gold nanocapsules were loaded with hydrophobic or hydrophilic drugs dissolved in a phase-change material (PCM) [139]. In this hybrid system,

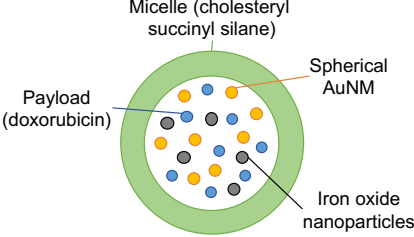
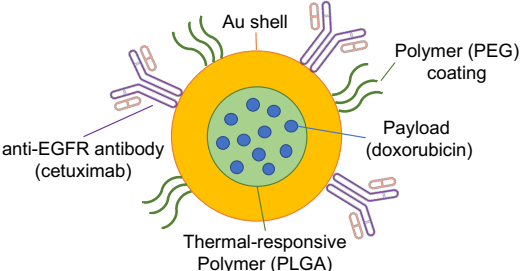

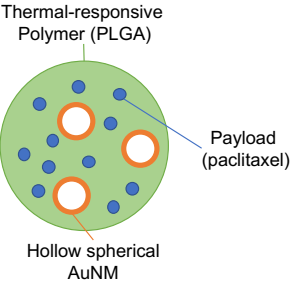
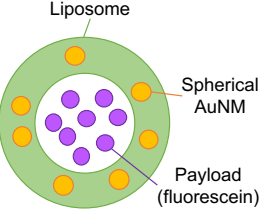
drug release is achieved when the PCM reaches the melting point, allowing the drug to diffuse out of the nanocapsules. HIFU was effective in controlling the drug release system by regulating the temperature and therefore the drug release profile.

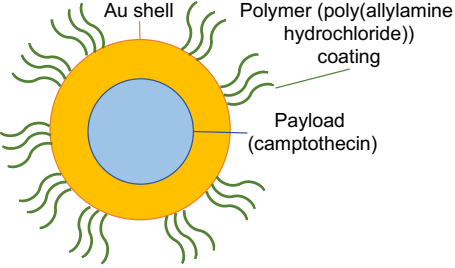
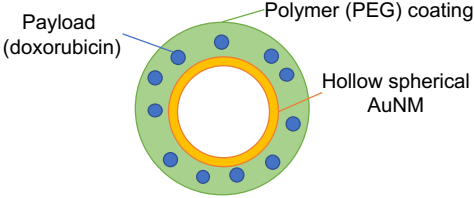
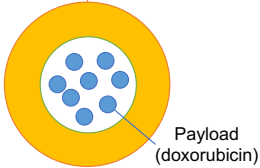
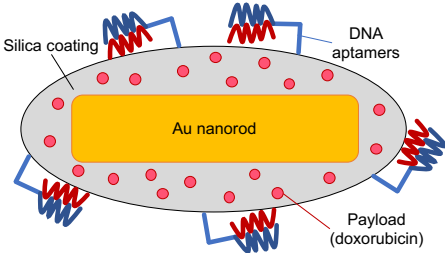
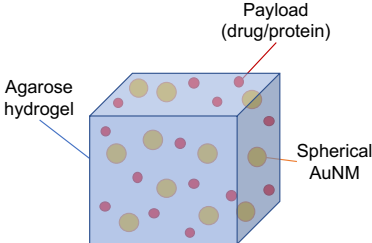
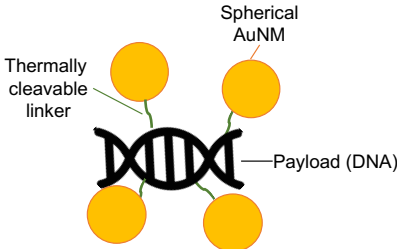
Radiotherapy

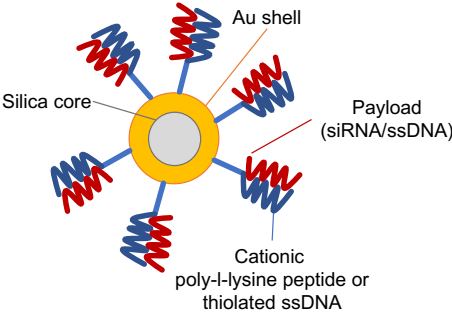
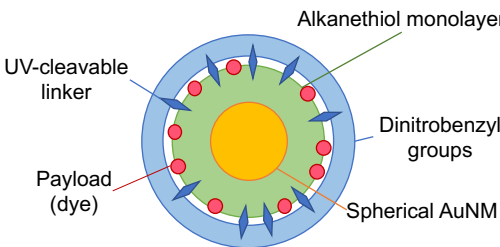
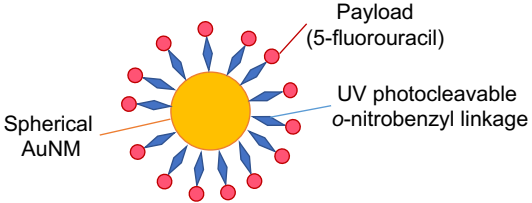
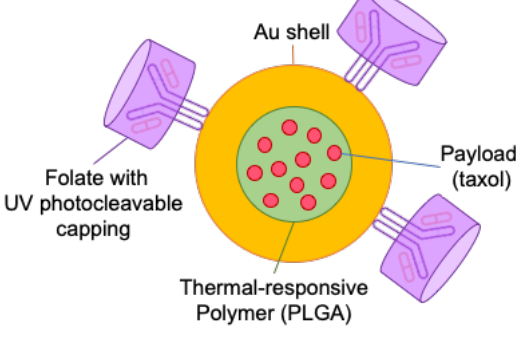
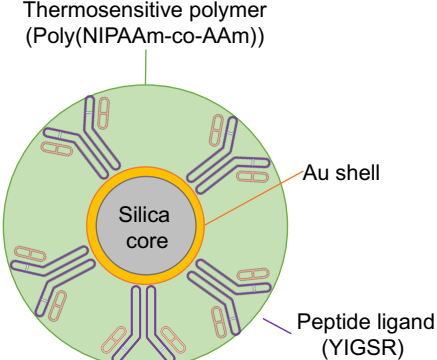
In radiotherapy, ionizing radiations (X-rays) are administered to tumor tissues to destroy the malignant cells directly or via the generation of free radicals. Owing to the strong X-ray absorption shown by AuNMs, the use of this nanomaterial in radiotherapy can lead to the enhancement of the local radiation dose while reducing other tissues getting radiated [70, 140-145]. Cui et al. demonstrated that the effect of radiosensitization by AuNMs is closely related to cellular uptake [146]. Similarly, AuNMs size has been demonstrated to play a key role in their ionizing properties: AuNMs with a size around 13 nm possess, for example, a superior radioactive disruption capability as compared to nanoparticles of other sizes [147]. Other than size, different AuNMs shapes also play an influence on their efficiency in radiotherapy, with spherical nanoparticles being the best ionizing agent to use [148]. Finally, various imaging modalities (e.g., PTT) and other therapeutic approaches can be coupled to radiotherapy when using AuNMs, to improve the therapeutic outcome [149-152].

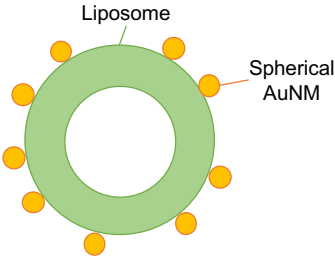
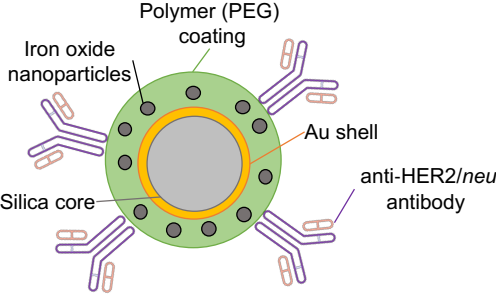
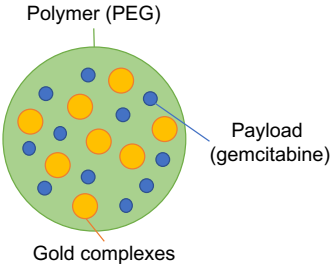
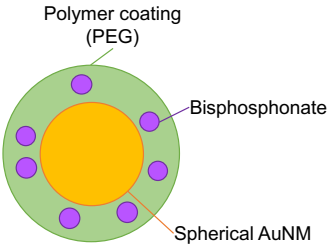
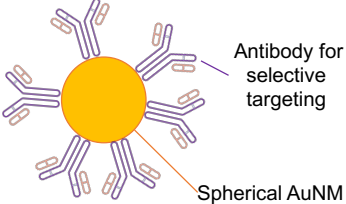
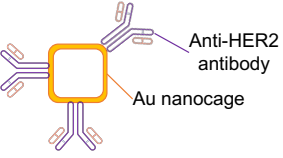
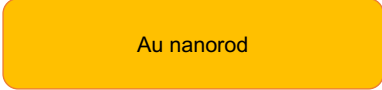
The AuNMs capability to amplify the X-ray local intensity can be used to develop smart drug delivery systems for selective drug activation [153]. Liu et al., for example, developed an X-ray-triggered nitrite release from nitroimidazole-functionalized AuNMs. Nitroimidazole is a pro-drug that, upon irradiation with X-rays, releases nitrite, a precursor of ROS, inducing the therapeutic effect against cancer cells [154]. The AuNMs radiosensitizing effect has also been exploited to trigger the release of an anti-cancer drug (doxorubicin, DOX) from DOX-conjugated DNA-coated AuNMs, resulting in a greater clonogenic cell kill compared to DOX-free DNA-coated AuNMs [155].

Table 2. Summary of the physically triggered AuNMs described in this review, for which the exogenous stimulus applied induces a structural transformation or modification that is essential for their therapeutic action. The AuNMs and their chemical components are not drawn to scale.

Therapeutic applications				
Physical stimulus type	Application	Strategy	AuNM structure	Ref
Laser light	Photo-responsive drug release	Thermal disruption of the compound coating the AuNMs surface or entrapped within the AuNMs, triggers the release of the therapeutic payload.		[63]
				[79]
				[80]
				[82]
				[85]

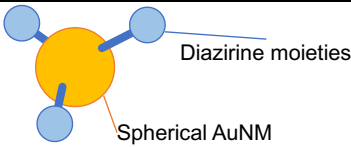

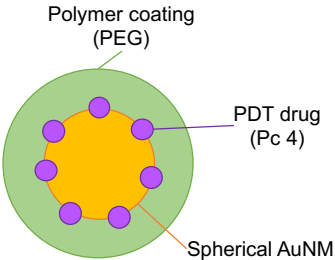
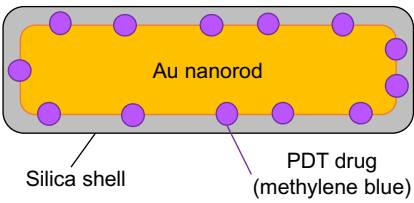
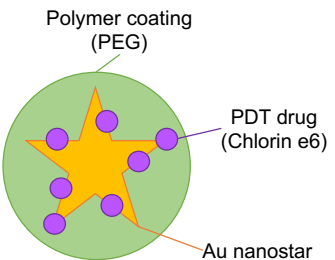
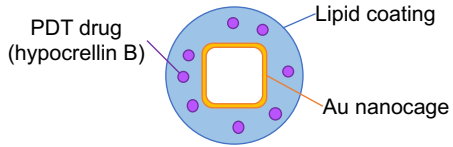
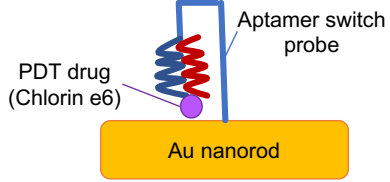
			 <p>Au shell Polymer (poly(allylamine hydrochloride)) coating Payload (camptothecin)</p>	[86]
			 <p>Payload (doxorubicin) Polymer (PEG) coating Hollow spherical AuNM</p>	[83,84,87]
			 <p>Hollow spherical AuNM Payload (doxorubicin)</p>	[89]
			 <p>Silica coating DNA aptamers Au nanorod Payload (doxorubicin)</p>	[90]
			 <p>Payload (drug/protein) Agarose hydrogel Spherical AuNM</p>	[91]
			 <p>Spherical AuNM Thermally cleavable linker Payload (DNA)</p>	[81]

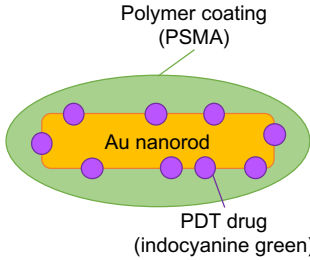
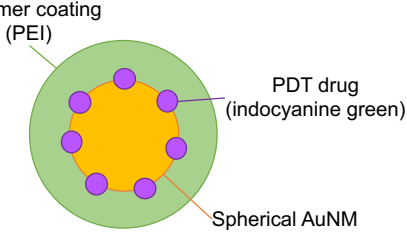
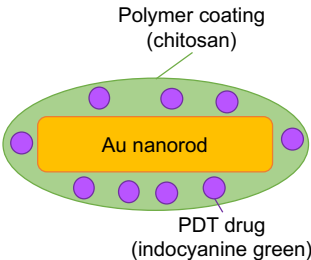
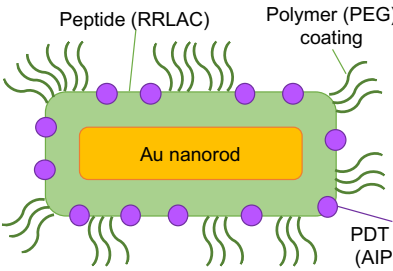
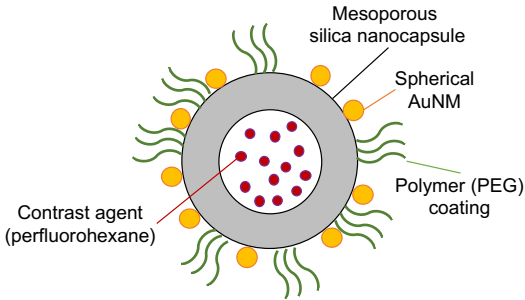
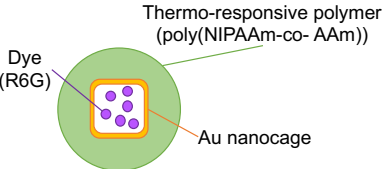
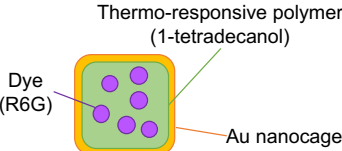
		<p>Thermal dehybridization allows the therapeutic payload (single-strand DNA, ssDNA, or siRNA) to be released.</p>	 <p>Au shell Silica core Payload (siRNA/ssDNA) Cationic poly-l-lysine peptide or thiolated ssDNA</p>	[92-96]
		<p>Photo-induced cleavage of the linkers conjugating AuNMs to the therapeutic payload.</p>	 <p>Alkanethiol monolayer UV-cleavable linker Payload (dye) Dinitrobenzyl groups Spherical AuNM</p>	[97]
			 <p>Payload (5-fluorouracil) UV photocleavable o-nitrobenzyl linkage Spherical AuNM</p>	[98]
	<p>Photo-responsive targeted therapy</p>	<p>Thermoresponsive polymers are used to mask a peptide ligand to control and reduce off-target effects.</p>	 <p>Au shell Folate with UV photocleavable capping Payload (taxol) Thermal-responsive Polymer (PLGA)</p>	[99]
			 <p>Thermosensitive polymer (Poly(NIPAAm-co-AAm)) Au shell Silica core Peptide ligand (YIGSR)</p>	[100]

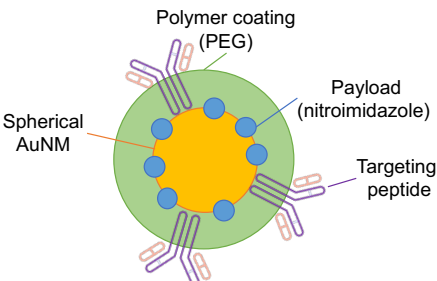
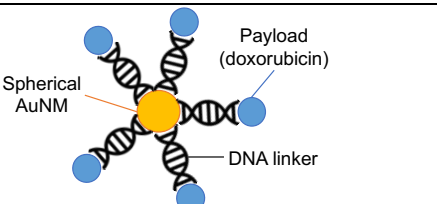
			 <p>Liposome Spherical AuNM</p>	[105]
			 <p>Polymer (PEG) coating Iron oxide nanoparticles Au shell anti-HER2/<i>neu</i> antibody Silica core</p>	[106]
			 <p>Polymer (PEG) Payload (gemcitabine) Gold complexes</p>	[103]
			 <p>Polymer coating (PEG) Bisphosphonate Spherical AuNM</p>	[104]
			 <p>Antibody for selective targeting Spherical AuNM</p>	[107]
			 <p>Anti-HER2 antibody Au nanocage</p>	[112]
			 <p>Au nanorod</p>	[42]

PTT

Light absorption and conversion into heat trigger cell death.

			 <p>Diazirine moieties Spherical AuNM</p>	[115]
	PDT	AuNMs are used as photosensitizing agents.	 <p>Au nanoechinus</p>	[116]
		AuNMs are used as carriers to deliver photosensitizers.	 <p>Polymer coating (PEG) PDT drug (Pc 4) Spherical AuNM</p>	[117]
			 <p>Au nanorod Silica shell PDT drug (methylene blue)</p>	[118]
			 <p>Polymer coating (PEG) PDT drug (Chlorin e6) Au nanostar</p>	[119]
			 <p>PDT drug (hypocrellin B) Lipid coating Au nanocage</p>	[120]
			 <p>PDT drug (Chlorin e6) Aptamer switch probe Au nanorod</p>	[121]

			 <p>Polymer coating (PSMA)</p> <p>Au nanorod</p> <p>PDT drug (indocyanine green)</p>  <p>Polymer coating (PEI)</p> <p>PDT drug (indocyanine green)</p> <p>Spherical AuNM</p>	[122]
			 <p>Polymer coating (chitosan)</p> <p>Au nanorod</p> <p>PDT drug (indocyanine green)</p>	[123]
			 <p>Peptide (RRLAC)</p> <p>Polymer (PEG) coating</p> <p>Au nanorod</p> <p>PDT drug (AIPcS₄)</p>	[124]
Ultrasound waves	High-intensity focused ultrasound therapy (HIFU)	AuNMs are used as HIFU agents, acting both as cavitation nuclei and promoting the collapse of the microbubbles within biological tissues.	 <p>Mesoporous silica nanocapsule</p> <p>Spherical AuNM</p> <p>Polymer (PEG) coating</p> <p>Contrast agent (perfluorohexane)</p>	[137]
			 <p>Dye (R6G)</p> <p>Thermo-responsive polymer (poly(NIPAAm-co-AAm))</p> <p>Au nanocage</p>	[138]
			 <p>Dye (R6G)</p> <p>Thermo-responsive polymer (1-tetradecanol)</p> <p>Au nanocage</p>	[139]

X-rays	Radiotherapy	AuNMs release a payload upon stimulation with X-rays.		[154]
				[155]

Diagnostic applications

AuNMs activated by ionizing and non-ionizing external physical stimuli can generate various signals, including light, Raman or Rayleigh scattering and ultrasonic waves, that are useful for diagnostic purposes. Additionally, owing to their tuneable surface functionalities and light absorption/scattering, and enhanced extinction coefficient as compared to most common dyes, AuNMs ensure both the selective targeting of the analyte/tissue of interest and a strong detecting signal. Wilhem et al. recently reviewed the state of the art in this field [156]. To the best of the authors' knowledge, however, AuNMs modifying their structure upon external physical stimuli to exert their diagnostic function, are less frequent than those designed for therapeutic applications. To date, AuNMs belonging to this specific category have been used in diagnostics only as optical biosensors and photoacoustic imaging (PAI) contrast agents. These AuNMs are described in more detail below and a schematic summary is presented in Table 3. The use of this type of AuNMs as a contrast agent for dark-field microscopy (iDFM), Raman imaging, photoacoustic tomography (PAT), MRI, and CT, are also interesting applications in the diagnostic field, but are still to be explored by scientists.

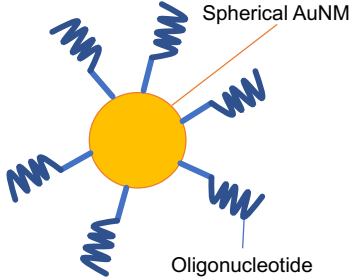
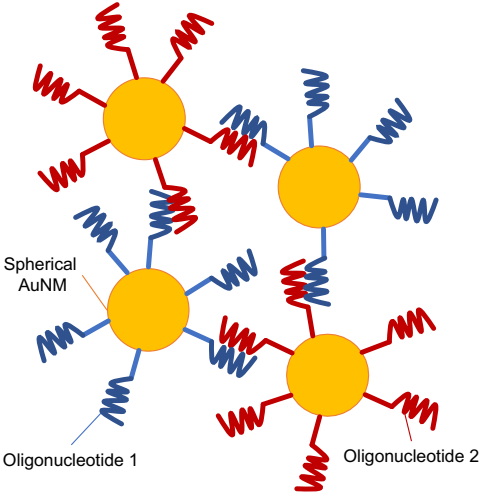
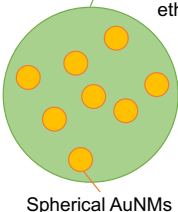
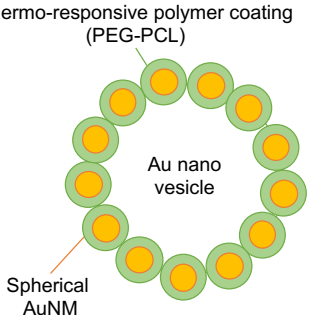
Optical biosensors

A typical AuNMs-based biosensor monitors the frequency shift in LSPR resonance [157]. Thus, the simplest implementation of LSPR sensing is the use of AuNMs as a functional

component of colorimetric assays both in liquid (homophase methods) and solid (dot immunoassay and immunochromatography) phase [1]. In these assays, AuNMs are conjugated with a bioactive moiety capable of binding with high-affinity specific analytes present in a solution [157, 158], such as biomolecules (e.g., proteins[159] or toxins[160]), small molecules (oligonucleotides [161]), ions (e.g., selenium [162]), or diseased cells (e.g., acute leukemia cells [163]). AuNMs-based optical sensors can also be used to detect specific antigens within the cellular compartments [164, 165]. In the presence of the analyte, the AuNMs-containing solution changes its optical/plasmon absorption (i.e., its color). These changes in the absorption spectrum can be easily detected either visually or spectrophotometrically, reaching a much lower limit of detection than conventional dyes. For example, conjugates of AuNMs-oligonucleotides are of great interest in the detection of DNA hybridization in the diagnosis of pathogenic and genetic diseases without the need of special instrumentation [1].

Thermoresponsive AuNMs are particularly useful in developing optical multiple aptasensors, such as DNA-aptamer-based colorimetric assays that can detect several targets at the same time in the liquid phase. In this instance, more than one class of aptamer (i.e., single-strain DNAs and oligopeptides with high binding affinity and selectivity for target molecules) are used to functionalize the surface of the AuNMs. The detection approach is based on the likelihood that the target DNA molecules

Table 3. Summary of the physically triggered gold nanomaterials described in this review, for which the exogeneous stimulus applied induces a structural transformation or modification that is essential for their diagnostic action. The AuNMs and their chemical components are not drawn to scale.

Diagnostic applications				
Stimulus type	Application	Strategy	AuNMs structure	Ref
Temperature	Optical multiplex biosensors	Target DNA molecules with different melting temperatures can be distinguished within the assay by AuNMs disassociation based on temperature.	 <p>Spherical AuNM</p> <p>Oligonucleotide</p>	[166, 167]
			 <p>Spherical AuNM</p> <p>Oligonucleotide 1</p> <p>Oligonucleotide 2</p>	[168]
		Thermoresponsive AuNMs are used as components of colorimetric assays to detect several targets at the same time.	 <p>Thermo-responsive polymer (poly(N-isopropylacrylamide-co-2-(dimethylamino)-ethylmethacrylate))</p> <p>Spherical AuNMs</p>	[169]
Laser light	Photoacoustic imaging (PAI)	AuNMs act as a contrast agent.	 <p>Thermo-responsive polymer coating (PEG-PCL)</p> <p>Au nano vesicle</p> <p>Spherical AuNM</p>	[175]

have different melting temperatures, and therefore they can be distinguished within the assay by AuNMs disassociation based on temperature [166-168].

Similarly, a thermoresponsive AuNMs biosensor capable of distinguishing between homocysteine and cysteine has been reported [169]. The assay used AuNMs conjugated with thermoresponsive copolymers. Such conjugation induced the assembly of the AuNMs in a liquid phase, changing the solution color from red to bluish purple. Heating the solution induced AuNMs disassembly in the presence of cysteine, but not in the presence of homocysteine, which inhibited the process.

Photoacoustic imaging (PAI)

PAI is based on the sample irradiation with a laser, by which the energy of light absorbed by AuNMs, used as exogenous contrast agents [170-172], leads to the generation of waves in an ultrasonic range that are then converted into an image. Compared to MRI and CT, PAI offers

various advantages including the use of non-ionizing irradiation, good spatial (5 μm -1 mm) and temporal (s-min) resolution, and improved sensitivity [173]. For example, AuNMs have been injected on-site into the tumor tissue, where they specifically bound to the cancer cells, helping to identify the malignant tissue and guiding the surgeons for precision treatment [174]. In PAI, larger AuNMs are preferred because of their higher scattering efficiency. Thus, multimodal gold nanovesicle, formed by the clustering of polymer-coated AuNMs, have been developed for simultaneous PAI and PTT [175]. Such clustering induces a shift in the absorption of AuNMs towards the NIR spectral region and a subsequent increase in PAI signal and image contrast, thus allowing to easily distinguish the tumor mass within the organ. Irradiation causes declustering of the gold nanovesicles, and the single AuNM components can be easily cleared from the body, avoiding systemic side effects.

Conclusions

In the last few decades, the growing incidence of chronic diseases associated with the increasing population aging, has triggered the need for more effective and localized therapies as well as efficient diagnostic tools for early disease detection [156]. AuNMs and alloy/hybrid AuNMs have been widely investigated to address this need. In this context, various synthetic strategies have been used to produce gold nanomedicines that enable target-specific treatments and robust diagnostic agents. The gold nanomaterials reviewed herein constitute the next-generation technology: they are AuNMs or hybrid/alloy AuNMs that are activated by external physical stimuli, where the therapeutic and/or diagnostic action is a result of the structural modification of the nanomaterial itself. Although this technology offers many advantages, such as a complete temporal and spatial control over the mode of action, these gold-based platforms are still at their infancy and they have not yet reached clinical phases. To increase the translation rate of this promising technology, the authors believe that future research efforts should focus on meeting clinical attributes, as recently discussed by Richardson et al. [176]. Such attributes should be investigated at the pre-clinical stages, to prove the existence of an advantageous risk-benefit ratio for patients. The adoption of the MIRIBEL (Minimum Information Reporting in Bio-Nano Experimental Literature) guidelines for published accounts of bio-nano research can support this activity [177]. On the other hand, technological requirements should also be satisfied, as pointed out over the years by experts in the field through many recent papers and reviews [178, 179], to ensure reproducibility and comparability [180]. We believe, within this frameset, an acceleration on the physically triggered gold nanomaterial products to clinics is foreseeable, with great achievements for the quality of life of patients.

Acknowledgements

This study has been funded, in parts, by the Center for Alternatives to Animal Testing Award (CAAT Project #2018-17). Also, the research leading to these results has received funding from the European Union's HORIZON 2020 Framework Programme under grant agreement NO. 760928.

The funding sponsors were not involved in the collection, analysis and interpretation of the information reviewed; in the writing of the manuscript; or in the decision to submit the article for publication.

Conflict of Interests

The authors declare no conflicts of interest. For signed statements, please contact the journal office: editor@precisionnanomedicine.com

Quote this article as: Movia D, Benhaddada M, Spadavecchia J, Prina-Mello A, Latest advances in combining gold nanomaterials with physical stimuli towards new responsive therapeutic and diagnostic strategies, *Precis. Nanomed.* 2020;3(2):495-524, <https://doi.org/10.33218/001c.12650>

Bibliography

- [1] H. Daraee, A. Eatemadi, E. Abbasi, S. Fekri Aval, M. Kouhi, and A. Akbarzadeh, "Application of gold nanoparticles in biomedical and drug delivery," *Artif Cells Nanomed Biotechnol*, vol. 44, no. 1, pp. 410-22, 2016, doi: 10.3109/21691401.2014.955107.
- [2] A. C. Anselmo and S. Mitragotri, "Nanoparticles in the clinic," (in eng), *Bioeng Transl Med*, vol. 1, no. 1, pp. 10-29, Mar 2016, doi: 10.1002/btm2.10003.
- [3] A. Gharatape and R. Salehi, "Recent progress in theranostic applications of hybrid gold nanoparticles," (in English), *Eur J Med Chem*, vol. 138, pp. 221-233, Sep 29 2017, doi: 10.1016/j.ejmech.2017.06.034.
- [4] J. Guo, K. Rahme, Y. He, L. L. Li, J. D. Holmes, and C. M. O'Driscoll, "Gold nanoparticles enlighten the future of cancer theranostics," (in English), *Int J Nanomed*, vol. 12, pp. 6131-6152, 2017, doi: 10.2147/IJN.S140772.
- [5] L. A. Dykman and N. G. Khlebtsov, "Multifunctional gold-based nanocomposites for theranostics," *Biomaterials*, vol. 108, pp. 13-34, Nov 2016, doi: 10.1016/j.biomaterials.2016.08.040.
- [6] C. Xu et al., "Au-Fe₃O₄ dumbbell nanoparticles as dual-functional probes," (in eng), *Angewandte Chemie (International ed. in English)*, vol. 47, no. 1, pp. 173-6, 2008, doi: 10.1002/anie.200704392.
- [7] J. S. Choi, Y. W. Jun, S. I. Yeon, H. C. Kim, J. S. Shin, and J. Cheon, "Biocompatible heterostructured nanoparticles for multimodal biological detection," (in eng), *Journal of the American Chemical Society*, vol. 128, no. 50, pp. 15982-3, Dec 20 2006, doi: 10.1021/ja066547g.
- [8] I. Capek, "Polymer decorated gold nanoparticles in nanomedicine conjugates," *Adv Colloid Interface Sci*, vol. 249, pp. 386-399, Nov 2017, doi: 10.1016/j.cis.2017.01.007.
- [9] P. Yadav, S. P. Singh, A. K. Rengan, A. Shanavas, and R. Srivastava, "Gold laced bio-macromolecules for theranostic application," *Int J Biol Macromol*, vol. 110, pp. 39-53, Apr 15 2018, doi: 10.1016/j.ijbiomac.2017.10.124.
- [10] F. Farjadian, A. Ghasemi, O. Gohari, A. Roointan, M. Karimi, and M. R. Hamblin, "Nanopharmaceuticals and nanomedicines currently on the market: challenges and opportunities," (in English), *Nanomedicine-Uk*, vol. 14, no. 1, pp. 93-126, Jan 2019, doi: 10.2217/nmm-2018-0120.
- [11] P. Singh, S. Pandit, V. Mokkalapati, A. Garg, V. Ravikumar, and I. Mijakovic, "Gold Nanoparticles in Diagnostics and Therapeutics for Human Cancer," (in English), *Int J Mol Sci*, vol. 19, no. 7, Jul 6 2018, doi: 10.3390/ijms19071979.
- [12] D. Bobo, K. J. Robinson, J. Islam, K. J. Thurecht, and S. R. Corrie, "Nanoparticle-Based Medicines: A Review of FDA-Approved Materials and Clinical Trials to Date," (in eng), *Pharm Res*, vol. 33, no. 10, pp. 2373-87, Oct 2016, doi: 10.1007/s11095-016-1958-5.
- [13] FDA. <https://www.fda.gov/vaccines-blood-biologics/approved-blood-products/uni-gold-recombinant-hiv-12> (accessed April, 2020).

- [14] FDA. <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfRL/rl.cfm?lid=423557&lpcd=LCX> (accessed April, 2020).
- [15] S. Tran, P. J. DeGiovanni, B. Piel, and P. Rai, "Cancer nanomedicine: a review of recent success in drug delivery," *Clin Transl Med*, vol. 6, no. 1, p. 44, Dec 11 2017, doi: 10.1186/s40169-017-0175-0.
- [16] L. Tian, L. Lu, Y. Qiao, S. Ravi, F. Salatan, and M. P. Melancon, "Stimuli-Responsive Gold Nanoparticles for Cancer Diagnosis and Therapy," (in English), *J Funct Biomater*, vol. 7, no. 3, Jul 21 2016, doi: 10.3390/jfb7020019.
- [17] F. Li et al., "Stimuli-responsive nano-assemblies for remotely controlled drug delivery," *Journal of Controlled Release*, 2020/04/08/ 2020, doi: 10.1016/j.jconrel.2020.03.051.
- [18] W. Gao, J. M. Chan, and O. C. Farokhzad, "pH-Responsive nanoparticles for drug delivery," *Mol Pharm*, vol. 7, no. 6, pp. 1913-20, Dec 6 2010, doi: 10.1021/mp100253e.
- [19] J. Yang, Y. Duan, X. Zhang, Y. Wang, and A. Yu, "Modulating the cellular microenvironment with disulfide-containing nanoparticles as an auxiliary cancer treatment strategy," (in English), *J Mater Chem B*, 10.1039/C6TB00847J vol. 4, no. 22, pp. 3868-3873, 2016, doi: 10.1039/c6tb00847j.
- [20] J. Spadavecchia et al., "Targeted polyethylene glycol gold nanoparticles for the treatment of pancreatic cancer: from synthesis to proof-of-concept in vitro studies," *Int J Nanomed*, vol. 11, pp. 791-822, 2016, doi: 10.2147/IJN.S97476.
- [21] R. J. Stafford, A. Shetty, A. M. Elliott, J. A. Schwartz, G. P. Goodrich, and J. D. Hazle, "MR temperature imaging of nanoshell mediated laser ablation," (in eng), *International journal of hyperthermia : the official journal of European Society for Hyperthermic Oncology, North American Hyperthermia Group*, vol. 27, no. 8, pp. 782-90, 2011, doi: 10.3109/02656736.2011.614671.
- [22] D. P. O'Neal, L. R. Hirsch, N. J. Halas, J. D. Payne, and J. L. West, "Photo-thermal tumor ablation in mice using near infrared-absorbing nanoparticles," (in eng), *Cancer letters*, vol. 209, no. 2, pp. 171-6, Jun 25 2004, doi: 10.1016/j.canlet.2004.02.004.
- [23] A. M. Gobin, M. H. Lee, N. J. Halas, W. D. James, R. A. Drezek, and J. L. West, "Near-infrared resonant nanoshells for combined optical imaging and photothermal cancer therapy," (in eng), *Nano letters*, vol. 7, no. 7, pp. 1929-34, Jul 2007, doi: 10.1021/nl070610y.
- [24] E. S. Day et al., "Nanoshell-mediated photothermal therapy improves survival in a murine glioma model," *J Neurooncol*, journal article vol. 104, no. 1, pp. 55-63, Aug 2011, doi: 10.1007/s11060-010-0470-8.
- [25] J. A. Schwartz et al., "Feasibility study of particle-assisted laser ablation of brain tumors in orthotopic canine model," (in eng), *Cancer research*, vol. 69, no. 4, pp. 1659-67, Feb 15 2009, doi: 10.1158/0008-5472.CAN-08-2535.
- [26] S. C. Gad, K. L. Sharp, C. Montgomery, J. D. Payne, and G. P. Goodrich, "Evaluation of the toxicity of intravenous delivery of auroshell particles (gold-silica nanoshells)," (in eng), *International journal of toxicology*, vol. 31, no. 6, pp. 584-94, Nov-Dec 2012, doi: 10.1177/1091581812465969.
- [27] J. M. Stern, V. V. Kibanov Solomonov, E. Sazykina, J. A. Schwartz, S. C. Gad, and G. P. Goodrich, "Initial Evaluation of the Safety of Nanoshell-Directed Photothermal Therapy in the Treatment of Prostate Disease," *International journal of toxicology*, vol. 35, no. 1, pp. 38-46, Jan-Feb 2016, doi: 10.1177/1091581815600170.
- [28] M. L. Li, J. C. Wang, J. A. Schwartz, K. L. Gill-Sharp, G. Stoica, and L. V. Wang, "In-vivo photoacoustic microscopy of nanoshell extravasation from solid tumor vasculature," (in eng), *Journal of biomedical optics*, vol. 14, no. 1, p. 010507, Jan-Feb 2009, doi: 10.1117/1.3081556.
- [29] L. Bregoli, D. Movia, J. D. Gavigan-Imedio, J. Lysaght, J. Reynolds, and A. Prina-Mello, "Nanomedicine applied to translational oncology: A future perspective on cancer treatment," (in eng), *Nanomedicine*, vol. 12, no. 1, pp. 81-103, Jan 2016, doi: 10.1016/j.nano.2015.08.006.

- [30] H. Maeda, "Macromolecular therapeutics in cancer treatment: the EPR effect and beyond," *Journal of controlled release : official journal of the Controlled Release Society*, vol. 164, no. 2, pp. 138-44, Dec 10 2012, doi: 10.1016/j.jconrel.2012.04.038.
- [31] I. A. Khawar, J. H. Kim, and H. J. Kuh, "Improving drug delivery to solid tumors: priming the tumor microenvironment," (in eng), *Journal of controlled release : official journal of the Controlled Release Society*, vol. 201, no. 1873-4995 (Electronic), pp. 78-89, Mar 10 2015, doi: 10.1016/j.jconrel.2014.12.018.
- [32] O. Tredan, C. M. Galmarini, K. Patel, and I. F. Tannock, "Drug resistance and the solid tumor microenvironment," (in eng), *J Natl Cancer Inst*, vol. 99, no. 19, pp. 1441-54, Oct 3 2007, doi: 10.1093/jnci/djm135.
- [33] T. Stylianopoulos and R. K. Jain, "Design considerations for nanotherapeutics in oncology," *Nanomedicine*, vol. 11, no. 8, pp. 1893-907, Nov 2015, doi: 10.1016/j.nano.2015.07.015.
- [34] J. M. Stern, J. Stanfield, Y. Lotan, S. Park, J. T. Hsieh, and J. A. Cadegdu, "Efficacy of laser-activated gold nanoshells in ablating prostate cancer cells in vitro," (in eng), *Journal of endourology*, vol. 21, no. 8, pp. 939-43, Aug 2007, doi: 10.1089/end.2007.0437.
- [35] J. M. Stern, J. Stanfield, W. Kabbani, J. T. Hsieh, and J. A. Cadegdu, "Selective prostate cancer thermal ablation with laser activated gold nanoshells," (in eng), *The Journal of urology*, vol. 179, no. 2, pp. 748-53, Feb 2008, doi: 10.1016/j.juro.2007.09.018.
- [36] F. F. Sahle, M. Gulfam, and T. L. Lowe, "Design strategies for physical-stimuli-responsive programmable nanotherapeutics," (in eng), *Drug Discov Today*, vol. 23, no. 5, pp. 992-1006, May 2018, doi: 10.1016/j.drudis.2018.04.003.
- [37] P. K. Jain, "Gold Nanoparticles for Physics, Chemistry, and Biology. Edited by Catherine Louis and Olivier Pluchery," *Angewandte Chemie International Edition*, vol. 53, no. 5, pp. 1197-1197, 2014, doi: 10.1002/anie.201309807.
- [38] L. Jing-Liang and G. Min, "Gold-Nanoparticle-Enhanced Cancer Photothermal Therapy," (in English), *Ieee J Sel Top Quant*, vol. 16, no. 4, pp. 989-996, Jul-Aug 2010, doi: 10.1109/jstqe.2009.2030340.
- [39] Z. Qin and J. C. Bischof, "Thermophysical and biological responses of gold nanoparticle laser heating," *Chem Soc Rev*, vol. 41, no. 3, pp. 1191-217, Feb 7 2012, doi: 10.1039/c1cs15184c.
- [40] X. Huang and M. A. El-Sayed, "Gold nanoparticles: Optical properties and implementations in cancer diagnosis and photothermal therapy," *Journal of Advanced Research*, vol. 1, no. 1, pp. 13-28, 2010/01/01/ 2010, doi: 10.1016/j.jare.2010.02.002.
- [41] K. Jiang, D. A. Smith, and A. Pinchuk, "Size-Dependent Photothermal Conversion Efficiencies of Plasmonically Heated Gold Nanoparticles," *The Journal of Physical Chemistry C*, vol. 117, no. 51, pp. 27073-27080, 2013/12/27 2013, doi: 10.1021/jp409067h.
- [42] Z. Qin et al., "Quantitative Comparison of Photothermal Heat Generation between Gold Nanospheres and Nanorods," (in eng), *Sci Rep*, vol. 6, p. 29836, Jul 21 2016, doi: 10.1038/srep29836.
- [43] B. Taghizadeh et al., "Classification of stimuli-responsive polymers as anticancer drug delivery systems," (in eng), *Drug Deliv*, vol. 22, no. 2, pp. 145-55, Feb 2015, doi: 10.3109/10717544.2014.887157.
- [44] T. Kawano et al., "Stabilizing of plasmid DNA in vivo by PEG-modified cationic gold nanoparticles and the gene expression assisted with electrical pulses," (in eng), *Journal of controlled release : official journal of the Controlled Release Society*, vol. 111, no. 3, pp. 382-9, Apr 10 2006, doi: 10.1016/j.jconrel.2005.12.022.
- [45] S. J. Corr et al., "Citrate-capped gold nanoparticle electrophoretic heat production in response to a time-varying radiofrequency electric-field," (in English), *J Phys Chem C*, vol. 116, no. 45, pp. 24380-24389, Nov 15 2012, doi: 10.1021/jp309053z.
- [46] S. M. Amini, S. Kharrazi, and M. R. Jaafari, "Radio frequency hyperthermia of cancerous cells with gold nanoclusters: an in vitro investigation," (in English), *Gold Bull*, vol. 50, no. 1, pp. 43-50, Mar 2017, doi: 10.1007/s13404-016-0192-6.

- [47] C. B. Collins, R. S. McCoy, B. J. Ackerson, G. J. Collins, and C. J. Ackerson, "Radiofrequency heating pathways for gold nanoparticles," (in eng), *Nanoscale*, vol. 6, no. 15, pp. 8459-72, Aug 7 2014, doi: 10.1039/c4nr00464g.
- [48] G. Canavese et al., "Nanoparticle-assisted ultrasound: A special focus on sonodynamic therapy against cancer," (in English), *Chem Eng J*, vol. 340, pp. 155-172, May 15 2018, doi: 10.1016/j.cej.2018.01.060.
- [49] A. Sazgarnia, A. Shanei, N. T. Meibodi, H. Eshghi, and H. Nassirli, "A novel nanosensitizer for sonodynamic therapy: in vivo study on a colon tumor model," (in English), *J Ultras Med*, vol. 30, no. 10, pp. 1321-9, Oct 2011, doi: 10.7863/jum.2011.30.10.1321.
- [50] J. L. Jiménez Pérez, A. Cruz-Orea, E. Ramón-Gallegos, R. Gutierrez Fuentes, and J. F. Sanchez Ramirez, "Photoacoustic Spectroscopy to determine in vitro the non radiative relaxation time of protoporphyrin IX solution containing gold metallic nanoparticles," *The European Physical Journal Special Topics*, vol. 153, no. 1, pp. 353-356, 2008/01/01 2008, doi: 10.1140/epjst/e2008-00460-2.
- [51] C. Brazzale et al., "Enhanced selective sonosensitizing efficacy of ultrasound-based anticancer treatment by targeted gold nanoparticles," (in English), *Nanomedicine-Uk*, vol. 11, no. 23, pp. 3053-3070, Dec 2016, doi: 10.2217/nmm-2016-0293.
- [52] M. Sadeghi-Goughari, S. Jeon, and H. J. Kwon, "Enhancing Thermal Effect of Focused Ultrasound Therapy Using Gold Nanoparticles," (in eng), *IEEE Trans Nanobioscience*, vol. 18, no. 4, pp. 661-668, Oct 2019, doi: 10.1109/TNB.2019.2937327.
- [53] J. R. McLaughlan, D. M. J. Cowell, and S. Freear, "Gold nanoparticle nucleated cavitation for enhanced high intensity focused ultrasound therapy," (in eng), *Phys Med Biol*, vol. 63, no. 1, p. 015004, Dec 14 2017, doi: 10.1088/1361-6560/aa97e9.
- [54] J. R. McLaughlan, "Controllable Nucleation of Cavitation from Plasmonic Gold Nanoparticles for Enhancing High Intensity Focused Ultrasound Applications," (in eng), *J Vis Exp*, no. 140, Oct 5 2018, doi: 10.3791/58045.
- [55] A. Shanei and A. Sazgarnia, "An overview of therapeutic applications of ultrasound based on synergetic effects with gold nanoparticles and laser excitation," (in eng), *Iran J Basic Med Sci*, vol. 22, no. 8, pp. 848-855, Aug 2019, doi: 10.22038/ijbms.2019.29584.7142.
- [56] M. Walter et al., "A unified view of ligand-protected gold clusters as superatom complexes," (in English), *P Natl Acad Sci USA*, vol. 105, no. 27, pp. 9157-62, Jul 8 2008, doi: 10.1073/pnas.0801001105.
- [57] R. S. McCoy, S. Choi, G. Collins, B. J. Ackerson, and C. J. Ackerson, "Superatom paramagnetism enables gold nanocluster heating in applied radiofrequency fields," (in English), *Acc Nano*, vol. 7, no. 3, pp. 2610-6, Mar 26 2013, doi: 10.1021/nn306015c.
- [58] M. A. Tofanelli and C. J. Ackerson, "Superatom electron configuration predicts thermal stability of Au₂₅(SR)₁₈ nanoclusters," (in English), *Journal of the American Chemical Society*, vol. 134, no. 41, pp. 16937-40, Oct 17 2012, doi: 10.1021/ja3072644.
- [59] M. Zhu et al., "Reversible switching of magnetism in thiolate-protected Au₂₅ superatoms," *Journal of the American Chemical Society*, vol. 131, no. 7, pp. 2490-2, Feb 25 2009, doi: 10.1021/ja809157f.
- [60] G. L. Nealon, B. Donnio, R. Greget, J. P. Kappler, E. Terazzi, and J. L. Gallani, "Magnetism in gold nanoparticles," *Nanoscale*, 10.1039/C2NR30640A vol. 4, no. 17, pp. 5244-58, Sep 7 2012, doi: 10.1039/c2nr30640a.
- [61] S. Trudel, "Unexpected magnetism in gold nanostructures: making gold even more attractive," *Gold Bull*, vol. 44, no. 1, pp. 3-13, 2011/03/01 2011, doi: 10.1007/s13404-010-0002-5.
- [62] S. M. Amini, "Gold nanostructures absorption capacities of various energy forms for thermal therapy applications," (in English), *J Therm Biol*, vol. 79, pp. 81-84, Jan 2019, doi: 10.1016/j.jtherbio.2018.12.007.

- [63] Y. Ma, X. Liang, S. Tong, G. Bao, Q. Ren, and Z. Dai, "Gold Nanoshell Nanomicelles for Potential Magnetic Resonance Imaging, Light-Triggered Drug Release, and Photothermal Therapy," (in English), *Adv Funct Mater*, vol. 23, no. 7, pp. 815-822, Feb 18 2013, doi: 10.1002/adfm.201201663.
- [64] N. S. Elbially et al., "Multifunctional magnetic-gold nanoparticles for efficient combined targeted drug delivery and interstitial photothermal therapy," *Int J Pharm*, vol. 554, pp. 256-263, Jan 10 2019, doi: 10.1016/j.ijpharm.2018.11.021.
- [65] L. Leon Felix et al., "Gold-decorated magnetic nanoparticles design for hyperthermia applications and as a potential platform for their surface-functionalization," *Sci Rep*, vol. 9, no. 1, p. 4185, Mar 12 2019, doi: 10.1038/s41598-019-40769-2.
- [66] M. V. Efremova et al., "Magnetite-Gold nanohybrids as ideal all-in-one platforms for theranostics," *Sci Rep*, vol. 8, no. 1, p. 11295, Jul 26 2018, doi: 10.1038/s41598-018-29618-w.
- [67] R. Hu et al., "Core-Shell Magnetic Gold Nanoparticles for Magnetic Field-Enhanced Radio-Photothermal Therapy in Cervical Cancer," (in eng), *Nanomaterials (Basel)*, vol. 7, no. 5, p. 111, May 11 2017, doi: 10.3390/nano7050111.
- [68] S. Moraes Silva, R. Tavallaie, L. Sandiford, R. D. Tilley, and J. J. Gooding, "Gold coated magnetic nanoparticles: from preparation to surface modification for analytical and biomedical applications," *Chem Commun*, 10.1039/C6CC03225G vol. 52, no. 48, pp. 7528-40, Jun 18 2016, doi: 10.1039/c6cc03225g.
- [69] W. Sun et al., "Red-Light-Controlled Release of Drug-Ru Complex Conjugates from Metallopolymer Micelles for Phototherapy in Hypoxic Tumor Environments," (in English), *Adv Funct Mater*, Article vol. 28, no. 39, Sep 26 2018, Art no. 1804227, doi: 10.1002/adfm.201804227.
- [70] S. Her, D. A. Jaffray, and C. Allen, "Gold nanoparticles for applications in cancer radiotherapy: Mechanisms and recent advancements," (in eng), *Advanced drug delivery reviews*, vol. 109, pp. 84-101, Jan 15 2017, doi: 10.1016/j.addr.2015.12.012.
- [71] N. M. Dimitriou et al., "Gold nanoparticles, radiations and the immune system: Current insights into the physical mechanisms and the biological interactions of this new alliance towards cancer therapy," (in English), *Pharmacol Therapeut*, vol. 178, pp. 1-17, Oct 2017, doi: 10.1016/j.pharmthera.2017.03.006.
- [72] J. Schuemann et al., "Roadmap to Clinical Use of Gold Nanoparticles for Radiation Sensitization," (in English), *Int J Radiat Oncol*, vol. 94, no. 1, pp. 189-205, Jan 1 2016, doi: 10.1016/j.ijrobp.2015.09.032.
- [73] L. E. Cole, R. D. Ross, J. M. Tilley, T. Vargo-Gogola, and R. K. Roeder, "Gold nanoparticles as contrast agents in x-ray imaging and computed tomography," *Nanomedicine-Uk*, vol. 10, no. 2, pp. 321-41, Jan 2015, doi: 10.2217/nnm.14.171.
- [74] B. Singhana, P. Slattery, A. Chen, M. Wallace, and M. P. Melancon, "Light-activatable gold nanoshells for drug delivery applications," (in English), *Aaps Pharmscitech*, vol. 15, no. 3, pp. 741-52, Jun 2014, doi: 10.1208/s12249-014-0097-8.
- [75] F. Y. Kong, J. W. Zhang, R. F. Li, Z. X. Wang, W. J. Wang, and W. Wang, "Unique Roles of Gold Nanoparticles in Drug Delivery, Targeting and Imaging Applications," (in eng), *Molecules (Basel, Switzerland)*, vol. 22, no. 9, Aug 31 2017, doi: 10.3390/molecules22091445.
- [76] A. Y. Rwei, W. Wang, and D. S. Kohane, "Photoreponsive nanoparticles for drug delivery," *Nano Today*, vol. 10, no. 4, pp. 451-467, Aug 1 2015, doi: 10.1016/j.nantod.2015.06.004.
- [77] H. Han, J. Y. Lee, and X. Lu, "Thermoresponsive nanoparticles + plasmonic nanoparticles = photoreponsive heterodimers: facile synthesis and sunlight-induced reversible clustering," (in English), *Chem Commun*, vol. 49, no. 55, pp. 6122-4, Jul 14 2013, doi: 10.1039/c3cc42273a.
- [78] X. Ai, J. Mu, and B. Xing, "Recent Advances of Light-Mediated Theranostics," (in eng), *Theranostics*, vol. 6, no. 13, pp. 2439-2457, 2016, doi: 10.7150/thno.16088.
- [79] J. Yang et al., "Smart drug-loaded polymer gold nanoshells for systemic and localized therapy of human epithelial cancer," (in English), *Adv Mater*, vol. 21, no. 43, pp. 4339-42, Nov 20 2009, doi: 10.1002/adma.200900334.

- [80] D. Wang et al., "Treatment of metastatic breast cancer by combination of chemotherapy and photothermal ablation using doxorubicin-loaded DNA wrapped gold nanorods," (in English), *Biomaterials*, vol. 35, no. 29, pp. 8374-84, Sep 2014, doi: 10.1016/j.biomaterials.2014.05.094.
- [81] G. Han et al., "Light-regulated release of DNA and its delivery to nuclei by means of photolabile gold nanoparticles," *Angewandte Chemie (International ed. in English)*, vol. 45, no. 19, pp. 3165-9, May 5 2006, doi: 10.1002/anie.200600214.
- [82] J. You, R. Shao, X. Wei, S. Gupta, and C. Li, "Near-infrared light triggers release of Paclitaxel from biodegradable microspheres: photothermal effect and enhanced antitumor activity," *Small*, vol. 6, no. 9, pp. 1022-31, May 7 2010, doi: 10.1002/sml.201000028.
- [83] J. You, G. Zhang, and C. Li, "Exceptionally high payload of doxorubicin in hollow gold nanospheres for near-infrared light-triggered drug release," (in eng), *Acs Nano*, vol. 4, no. 2, pp. 1033-41, Feb 23 2010, doi: 10.1021/nn901181c.
- [84] J. You et al., "Effective photothermal chemotherapy using doxorubicin-loaded gold nanospheres that target EphB4 receptors in tumors," (in English), *Cancer research*, vol. 72, no. 18, pp. 4777-86, Sep 15 2012, doi: 10.1158/0008-5472.CAN-12-1003.
- [85] H. L. Huang, P. H. Lu, H. C. Yang, G. D. Lee, H. R. Li, and K. C. Liao, "Fiber-optic triggered release of liposome in vivo: implication of personalized chemotherapy," (in English), *Int J Nanomed*, vol. 10, pp. 5171-84, 2015, doi: 10.2147/IJN.S85915.
- [86] W. Li et al., "Functional core/shell drug nanoparticles for highly effective synergistic cancer therapy," (in English), *Adv Healthc Mater*, vol. 3, no. 9, pp. 1475-85, Sep 2014, doi: 10.1002/adhm.201300577.
- [87] J. Park et al., "Multifunctional hollow gold nanoparticles designed for triple combination therapy and CT imaging," (in English), *Journal of controlled release : official journal of the Controlled Release Society*, vol. 207, pp. 77-85, Jun 10 2015, doi: 10.1016/j.jconrel.2015.04.007.
- [88] E. Y. Lukianova-Hleb, X. Ren, R. R. Sawant, X. Wu, V. P. Torchilin, and D. O. Lapotko, "On-demand intracellular amplification of chemoradiation with cancer-specific plasmonic nanobubbles," *Nat Med*, vol. 20, no. 7, pp. 778-784, Jul 2014, doi: 10.1038/nm.3484.
- [89] A. L. Tam et al., "Imaging Intratumoral Nanoparticle Uptake After Combining Nanoembolization with Various Ablative Therapies in Hepatic VX2 Rabbit Tumors," (in English), *J Biomed Nanotechnol*, vol. 12, no. 2, pp. 296-307, Feb 2016, doi: 10.1166/jbn.2016.2174.
- [90] X. Yang, X. Liu, Z. Liu, F. Pu, J. Ren, and X. Qu, "Near-infrared light-triggered, targeted drug delivery to cancer cells by aptamer gated nanovehicles," (in English), *Adv Mater*, vol. 24, no. 21, pp. 2890-5, Jun 5 2012, doi: 10.1002/adma.201104797.
- [91] J. S. Basuki et al., "Photo-Modulated Therapeutic Protein Release from a Hydrogel Depot Using Visible Light," (in English), *Angewandte Chemie (International ed. in English)*, vol. 56, no. 4, pp. 966-971, Jan 19 2017, doi: 10.1002/anie.201610618.
- [92] R. Huschka, A. Barhoumi, Q. Liu, J. A. Roth, L. Ji, and N. J. Halas, "Gene silencing by gold nanoshell-mediated delivery and laser-triggered release of antisense oligonucleotide and siRNA," (in eng), *Acs Nano*, vol. 6, no. 9, pp. 7681-91, Sep 25 2012, doi: 10.1021/nn301135w.
- [93] R. Huschka, O. Neumann, A. Barhoumi, and N. J. Halas, "Visualizing light-triggered release of molecules inside living cells," (in eng), *Nano letters*, vol. 10, no. 10, pp. 4117-4122, Oct 13 2010, doi: 10.1021/nl102293b.
- [94] R. S. Riley, M. N. Dang, M. M. Billingsley, B. Abraham, L. Gundlach, and E. S. Day, "Evaluating the Mechanisms of Light-Triggered siRNA Release from Nanoshells for Temporal Control Over Gene Regulation," (in eng), *Nano letters*, vol. 18, no. 6, pp. 3565-3570, Jun 13 2018, doi: 10.1021/acs.nanolett.8b00681.
- [95] A. M. Goodman, N. J. Hogan, S. Gottheim, C. Li, S. E. Clare, and N. J. Halas, "Understanding Resonant Light-Triggered DNA Release from Plasmonic Nanoparticles," (in eng), *Acs Nano*, vol. 11, no. 1, pp. 171-179, Jan 24 2017, doi: 10.1021/acsnano.6b06510.

[96] W. Lu et al., "Tumor site-specific silencing of NF-kappaB p65 by targeted hollow gold nanosphere-mediated photothermal transfection," *Cancer research*, vol. 70, no. 8, pp. 3177-88, Apr 15 2010, doi: 10.1158/0008-5472.CAN-09-3379.

[97] K. Sreejivungsa, N. Suchaichit, P. Moosophon, and A. Chompoosor, "Light-Regulated Release of Entrapped Drugs from Photoresponsive Gold Nanoparticles," (in English), *J Nanomater*, vol. 2016, pp. 1-7, 2016, Art no. 4964693, doi: 10.1155/2016/4964693.

[98] S. S. Agasti, A. Chompoosor, C. C. You, P. Ghosh, C. K. Kim, and V. M. Rotello, "Photoregulated release of caged anticancer drugs from gold nanoparticles," (in eng), *Journal of the American Chemical Society*, vol. 131, no. 16, pp. 5728-9, Apr 29 2009, doi: 10.1021/ja900591t.

[99] N. C. Fan, F. Y. Cheng, J. A. Ho, and C. S. Yeh, "Photocontrolled targeted drug delivery: photocaged biologically active folic acid as a light-responsive tumor-targeting molecule," (in eng), *Angewandte Chemie (International ed. in English)*, vol. 51, no. 35, pp. 8806-10, Aug 27 2012, doi: 10.1002/anie.201203339.

[100] A. Barhoumi, W. Wang, D. Zurakowski, R. S. Langer, and D. S. Kohane, "Photothermally targeted thermosensitive polymer-masked nanoparticles," (in eng), *Nano letters*, vol. 14, no. 7, pp. 3697-701, Jul 9 2014, doi: 10.1021/nl403733z.

[101] Y. Zhang et al., "Temperature-dependent cell death patterns induced by functionalized gold nanoparticle photothermal therapy in melanoma cells," *Sci Rep*, vol. 8, no. 1, p. 8720, Jun 7 2018, doi: 10.1038/s41598-018-26978-1.

[102] H. Norouzi, K. Khoshgard, and F. Akbarzadeh, "In vitro outlook of gold nanoparticles in photothermal therapy: a literature review," (in English), *Laser Med Sci*, vol. 33, no. 4, pp. 917-926, May 2018, doi: 10.1007/s10103-018-2467-z.

[103] H. Liu, P. Jiang, Z. Li, X. Li, N. Djaker, and J. Spadavecchia, "HIV-1 Tat Peptide-Gemcitabine Gold (III)-PEGylated Complex-Nanoflowers: A Sleek Thermosensitive Hybrid Nanocarrier as Prospective Anticancer," *Particle & Particle Systems Characterization*, vol. 35, no. 8, p. 1800082, 2018, doi: 10.1002/ppsc.201800082.

[104] M. Monteil et al., "Polyphosphonate ligands: From synthesis to design of hybrid PEGylated nanoparticles toward phototherapy studies," (in eng), *Journal of colloid and interface science*, vol. 513, pp. 205-213, Mar 1 2018, doi: 10.1016/j.jcis.2017.10.055.

[105] A. K. Rengan et al., "In vivo analysis of biodegradable liposome gold nanoparticles as efficient agents for photothermal therapy of cancer," (in English), *Nano letters*, vol. 15, no. 2, pp. 842-8, Feb 11 2015, doi: 10.1021/nl5045378.

[106] J. Kim et al., "Designed fabrication of multifunctional magnetic gold nanoshells and their application to magnetic resonance imaging and photothermal therapy," (in English), *Angewandte Chemie (International ed. in English)*, vol. 45, no. 46, pp. 7754-8, Nov 27 2006, doi: 10.1002/anie.200602471.

[107] C. M. Pitsillides, E. K. Joe, X. Wei, R. R. Anderson, and C. P. Lin, "Selective cell targeting with light-absorbing microparticles and nanoparticles," (in eng), *Biophys J*, vol. 84, no. 6, pp. 4023-32, Jun 2003, doi: 10.1016/S0006-3495(03)75128-5.

[108] S. Link and M. A. El-Sayed, "Shape and size dependence of radiative, non-radiative and photothermal properties of gold nanocrystals," (in English), *Int Rev Phys Chem*, vol. 19, no. 3, pp. 409-453, Jul-Sep 2000, doi: 10.1080/01442350050034180.

[109] R. S. Riley and E. S. Day, "Gold nanoparticle-mediated photothermal therapy: applications and opportunities for multimodal cancer treatment," *Wiley Interdiscip Rev Nanomed Nanobiotechnol*, vol. 9, no. 4, Jul 2017, doi: 10.1002/wnan.1449.

[110] N. S. Abadeer and C. J. Murphy, "Recent Progress in Cancer Thermal Therapy Using Gold Nanoparticles," (in English), *The Journal of Physical Chemistry C*, vol. 120, no. 9, pp. 4691-4716, Mar 10 2016, doi: 10.1021/acs.jpcc.5b11232.

[111] J. R. Cole, N. A. Mirin, M. W. Knight, G. P. Goodrich, and N. J. Halas, "Photothermal Efficiencies of Nanoshells and Nanorods for Clinical Therapeutic Applications," (in English), *The*

Journal of Physical Chemistry C, vol. 113, no. 28, pp. 12090-12094, Jul 16 2009, doi: 10.1021/jp9003592.

[112] L. Au, D. Zheng, F. Zhou, Z. Y. Li, X. Li, and Y. Xia, "A quantitative study on the photothermal effect of immuno gold nanocages targeted to breast cancer cells," (in English), *Acs Nano*, vol. 2, no. 8, pp. 1645-52, Aug 2008, doi: 10.1021/nn800370j.

[113] S. Hwang, J. Nam, S. Jung, J. Song, H. Doh, and S. Kim, "Gold nanoparticle-mediated photothermal therapy: current status and future perspective," (in eng), *Nanomedicine-Uk*, vol. 9, no. 13, pp. 2003-22, Sep 2014, doi: 10.2217/nnm.14.147.

[114] J. Mesiccek and K. Kuca, "Summary of numerical analyses for therapeutic uses of laser-activated gold nanoparticles," *International journal of hyperthermia : the official journal of European Society for Hyperthermic Oncology, North American Hyperthermia Group*, vol. 34, no. 8, pp. 1255-1264, Dec 2018, doi: 10.1080/02656736.2018.1440016.

[115] X. Cheng, R. Sun, L. Yin, Z. Chai, H. Shi, and M. Gao, "Light-Triggered Assembly of Gold Nanoparticles for Photothermal Therapy and Photoacoustic Imaging of Tumors In Vivo," *Adv Mater*, vol. 29, no. 6, p. 1604894, Feb 2017, doi: 10.1002/adma.201604894.

[116] P. Vijayaraghavan, C. H. Liu, R. Vankayala, C. S. Chiang, and K. C. Hwang, "Designing multi-branched gold nanoechinus for NIR light activated dual modal photodynamic and photothermal therapy in the second biological window," (in English), *Adv Mater*, vol. 26, no. 39, pp. 6689-95, Oct 22 2014, doi: 10.1002/adma.201400703.

[117] Y. Cheng, C. S. A, J. D. Meyers, I. Panagopoulos, B. Fei, and C. Burda, "Highly efficient drug delivery with gold nanoparticle vectors for in vivo photodynamic therapy of cancer," *Journal of the American Chemical Society*, vol. 130, no. 32, pp. 10643-7, Aug 13 2008, doi: 10.1021/ja801631c.

[118] S. H. Seo et al., "NIR-light-induced surface-enhanced Raman scattering for detection and photothermal/photodynamic therapy of cancer cells using methylene blue-embedded gold nanorod@SiO₂ nanocomposites," (in English), *Biomaterials*, vol. 35, no. 10, pp. 3309-18, Mar 2014, doi: 10.1016/j.biomaterials.2013.12.066.

[119] S. Wang et al., "Single continuous wave laser induced photodynamic/plasmonic photothermal therapy using photosensitizer-functionalized gold nanostars," (in English), *Adv Mater*, vol. 25, no. 22, pp. 3055-61, Jun 11 2013, doi: 10.1002/adma.201204623.

[120] L. Gao, J. Fei, J. Zhao, H. Li, Y. Cui, and J. Li, "Hypocrellin-loaded gold nanocages with high two-photon efficiency for photothermal/photodynamic cancer therapy in vitro," (in English), *Acs Nano*, vol. 6, no. 9, pp. 8030-40, Sep 25 2012, doi: 10.1021/nn302634m.

[121] J. Wang et al., "Assembly of aptamer switch probes and photosensitizer on gold nanorods for targeted photothermal and photodynamic cancer therapy," (in English), *Acs Nano*, vol. 6, no. 6, pp. 5070-7, Jun 26 2012, doi: 10.1021/nn300694v.

[122] W. S. Kuo et al., "Gold nanomaterials conjugated with indocyanine green for dual-modality photodynamic and photothermal therapy," *Biomaterials*, vol. 33, no. 11, pp. 3270-8, Apr 2012, doi: 10.1016/j.biomaterials.2012.01.035.

[123] R. Chen, X. Wang, X. Yao, X. Zheng, J. Wang, and X. Jiang, "Near-IR-triggered photothermal/photodynamic dual-modality therapy system via chitosan hybrid nanospheres," *Biomaterials*, vol. 34, no. 33, pp. 8314-22, Nov 2013, doi: 10.1016/j.biomaterials.2013.07.034.

[124] B. Jang, J. Y. Park, C. H. Tung, I. H. Kim, and Y. Choi, "Gold nanorod-photosensitizer complex for near-infrared fluorescence imaging and photodynamic/photothermal therapy in vivo," (in English), *Acs Nano*, vol. 5, no. 2, pp. 1086-94, Feb 22 2011, doi: 10.1021/nn102722z.

[125] N. Rapoport, A. M. Kennedy, J. E. Shea, C. L. Scaife, and K. H. Nam, "Ultrasonic nanotherapy of pancreatic cancer: lessons from ultrasound imaging," (in English), *Mol Pharm*, vol. 7, no. 1, pp. 22-31, Feb 1 2010, doi: 10.1021/mp900128x.

[126] R. B. Sequeiros, K. Joronen, G. Komar, and S. K. Koskinen, "High intensity focused ultrasound (HIFU) in tumor therapy," *Duodecim*, vol. 133, no. 2, pp. 143-9, 2017. [Online]. Available: <https://www.ncbi.nlm.nih.gov/pubmed/29205014>.

- [127] G. Ter Haar, "HIFU Tissue Ablation: Concept and Devices," *Adv Exp Med Biol*, vol. 880, pp. 3-20, 2016, doi: 10.1007/978-3-319-22536-4_1.
- [128] T. J. Dubinsky, C. Cuevas, M. K. Dighe, O. Kolokythas, and J. H. Hwang, "High-intensity focused ultrasound: current potential and oncologic applications," (in English), *Am J Roentgenol*, vol. 190, no. 1, pp. 191-9, Jan 2008, doi: 10.2214/AJR.07.2671.
- [129] A. Sazgarnia, A. Shanei, and M. M. Shanei, "Monitoring of transient cavitation induced by ultrasound and intense pulsed light in presence of gold nanoparticles," (in English), *Ultrason Sonochem*, vol. 21, no. 1, pp. 268-74, Jan 2014, doi: 10.1016/j.ultsonch.2013.07.008.
- [130] J. Xi et al., "Au nanoparticle-coated, PLGA-based hybrid capsules for combined ultrasound imaging and HIFU therapy," (in English), *J Mater Chem B*, vol. 3, no. 20, pp. 4213-4220, 2015, doi: 10.1039/c5tb00200a.
- [131] C. Tarapacki and R. Karshafian, "Enhancing laser therapy using PEGylated gold nanoparticles combined with ultrasound and microbubbles," (in English), *Ultrasonics*, vol. 57, pp. 36-43, Mar 2015, doi: 10.1016/j.ultras.2014.10.015.
- [132] S. Mo et al., "Increasing the density of nanomedicines improves their ultrasound-mediated delivery to tumours," (in English), *Journal of controlled release : official journal of the Controlled Release Society*, vol. 210, pp. 10-8, Jul 28 2015, doi: 10.1016/j.jconrel.2015.05.265.
- [133] A. Sazgarnia et al., "Therapeutic effects of acoustic cavitation in the presence of gold nanoparticles on a colon tumor model," *J Ultras Med*, vol. 32, no. 3, pp. 475-83, Mar 2013, doi: 10.7863/jum.2013.32.3.475.
- [134] O. K. Kosheleva, T. C. Lai, N. G. Chen, M. Hsiao, and C. H. Chen, "Selective killing of cancer cells by nanoparticle-assisted ultrasound," (in English), *J Nanobiotechnol*, vol. 14, no. 1, p. 46, Jun 14 2016, doi: 10.1186/s12951-016-0194-9.
- [135] A. Sazgarnia, A. Shanei, H. Eshghi, M. Hassanzadeh-Khayyat, H. Esmaily, and M. M. Shanei, "Detection of sonoluminescence signals in a gel phantom in the presence of Protoporphyrin IX conjugated to gold nanoparticles," (in English), *Ultrasonics*, vol. 53, no. 1, pp. 29-35, Jan 2013, doi: 10.1016/j.ultras.2012.03.009.
- [136] H. Ju, R. A. Roy, and T. W. Murray, "Gold nanoparticle targeted photoacoustic cavitation for potential deep tissue imaging and therapy," (in English), *Biomed Opt Express*, vol. 4, no. 1, pp. 66-76, Jan 1 2013, doi: 10.1364/BOE.4.000066.
- [137] X. Wang et al., "Au-nanoparticle coated mesoporous silica nanocapsule-based multifunctional platform for ultrasound mediated imaging, cytolysis and tumor ablation," *Biomaterials*, vol. 34, no. 8, pp. 2057-68, Mar 2013, doi: 10.1016/j.biomaterials.2012.11.044.
- [138] W. Li et al., "Gold nanocages covered with thermally-responsive polymers for controlled release by high-intensity focused ultrasound," (in English), *Nanoscale*, vol. 3, no. 4, pp. 1724-30, Apr 2011, doi: 10.1039/c0nr00932f.
- [139] G. D. Moon et al., "A new theranostic system based on gold nanocages and phase-change materials with unique features for photoacoustic imaging and controlled release," (in English), *Journal of the American Chemical Society*, vol. 133, no. 13, pp. 4762-5, Apr 6 2011, doi: 10.1021/ja200894u.
- [140] J. F. Hainfeld, D. N. Slatkin, and H. M. Smilowitz, "The use of gold nanoparticles to enhance radiotherapy in mice," (in English), *Phys Med Biol*, vol. 49, no. 18, pp. N309-15, Sep 21 2004, doi: 10.1088/0031-9155/49/18/n03.
- [141] S. Jain et al., "Cell-specific radiosensitization by gold nanoparticles at megavoltage radiation energies," (in English), *Int J Radiat Oncol*, vol. 79, no. 2, pp. 531-9, Feb 1 2011, doi: 10.1016/j.ijrobp.2010.08.044.
- [142] T. Kong et al., "Enhancement of radiation cytotoxicity in breast-cancer cells by localized attachment of gold nanoparticles," (in English), *Small*, vol. 4, no. 9, pp. 1537-43, Sep 2008, doi: 10.1002/sml.200700794.

- [143] Y. Zheng and L. Sanche, "Gold nanoparticles enhance DNA damage induced by anti-cancer drugs and radiation," (in English), *Radiat Res*, vol. 172, no. 1, pp. 114-9, Jul 2009, doi: 10.1667/RR1689.1.
- [144] L. Cui et al., "Significant Radiation Enhancement Effects by Gold Nanoparticles in Combination with Cisplatin in Triple Negative Breast Cancer Cells and Tumor Xenografts," (in English), *Radiat Res*, vol. 187, no. 2, pp. 147-160, Feb 2017, doi: 10.1667/RR14578.1.
- [145] E. S. Davidi et al., "Cisplatin-conjugated gold nanoparticles as a theranostic agent for head and neck cancer," (in English), *Head Neck-J Sci Spec*, vol. 40, no. 1, pp. 70-78, Jan 2018, doi: 10.1002/hed.24935.
- [146] L. Cui et al., "Hypoxia and cellular localization influence the radiosensitizing effect of gold nanoparticles (AuNPs) in breast cancer cells," (in English), *Radiat Res*, vol. 182, no. 5, pp. 475-88, Nov 2014, doi: 10.1667/RR13642.1.
- [147] Y. Dou et al., "Size-Tuning Ionization To Optimize Gold Nanoparticles for Simultaneous Enhanced CT Imaging and Radiotherapy," *Acs Nano*, vol. 10, no. 2, pp. 2536-48, Feb 23 2016, doi: 10.1021/acsnano.5b07473.
- [148] N. Ma et al., "Shape-Dependent Radiosensitization Effect of Gold Nanostructures in Cancer Radiotherapy: Comparison of Gold Nanoparticles, Nanospikes, and Nanorods," (in eng), *ACS applied materials & interfaces*, vol. 9, no. 15, pp. 13037-13048, Apr 19 2017, doi: 10.1021/acsmi.7b01112.
- [149] A. Popovtzer et al., "Actively targeted gold nanoparticles as novel radiosensitizer agents: an in vivo head and neck cancer model," (in eng), *Nanoscale*, vol. 8, no. 5, pp. 2678-85, Feb 7 2016, doi: 10.1039/c5nr07496g.
- [150] G. Liang, X. Jin, S. Zhang, and D. Xing, "RGD peptide-modified fluorescent gold nanoclusters as highly efficient tumor-targeted radiotherapy sensitizers," (in eng), *Biomaterials*, vol. 144, pp. 95-104, Nov 2017, doi: 10.1016/j.biomaterials.2017.08.017.
- [151] H. Zhou, Y. Zhang, G. Su, S. Zhai, and B. Yan, "Enhanced cancer cell killing by a targeting gold nanoconstruct with doxorubicin payload under X-ray irradiation," (in English), *Rsc Adv*, vol. 3, no. 44, pp. 21596-21603, 2013, doi: 10.1039/c3ra43838d.
- [152] E. Spyratou, M. Makropoulou, E. P. Efsthopoulos, A. G. Georgakilas, and L. Sihver, "Recent Advances in Cancer Therapy Based on Dual Mode Gold Nanoparticles," *Cancers (Basel)*, vol. 9, no. 12, Dec 19 2017, doi: 10.3390/cancers9120173.
- [153] N. M. S. Nagi, Y. A. M. Khair, and A. M. E. Abdalla, "Capacity of gold nanoparticles in cancer radiotherapy," (in English), *Jpn J Radiol*, vol. 35, no. 10, pp. 555-561, Oct 2017, doi: 10.1007/s11604-017-0671-6.
- [154] F. Liu, J. Lou, and D. Hristov, "X-Ray responsive nanoparticles with triggered release of nitrite, a precursor of reactive nitrogen species, for enhanced cancer radiosensitization," *Nanoscale*, 10.1039/C7NR04684G vol. 9, no. 38, pp. 14627-14634, Oct 5 2017, doi: 10.1039/c7nr04684g.
- [155] Z. B. Starkewolf, L. Miyachi, J. Wong, and T. Guo, "X-ray triggered release of doxorubicin from nanoparticle drug carriers for cancer therapy," *Chem Commun*, vol. 49, no. 25, pp. 2545-7, Mar 28 2013, doi: 10.1039/c3cc38100e.
- [156] C. Wilhelm, F. Gazeau, and A. K. A. Silva, "Physically-triggered nanosystems for therapy and diagnosis," (in eng), *Advanced drug delivery reviews*, vol. 138, pp. 1-2, Jan 1 2019, doi: 10.1016/j.addr.2019.03.001.
- [157] H. Aldewachi, T. Chalati, M. N. Woodroffe, N. Bricklebank, B. Sharrack, and P. Gardiner, "Gold nanoparticle-based colorimetric biosensors," (in English), *Nanoscale*, vol. 10, no. 1, pp. 18-33, Dec 21 2017, doi: 10.1039/c7nr06367a.
- [158] M. Sabela, S. Balme, M. Bechelany, J.-M. Janot, and K. Bisetty, "A Review of Gold and Silver Nanoparticle-Based Colorimetric Sensing Assays," (in English), *Adv Eng Mater*, vol. 19, no. 12, p. 1700270, Dec 2017, doi: 10.1002/adem.201700270.

- [159] C. S. Tsai, T. B. Yu, and C. T. Chen, "Gold nanoparticle-based competitive colorimetric assay for detection of protein-protein interactions," *Chem Commun*, 10.1039/B507237A no. 34, pp. 4273-5, Sep 14 2005, doi: 10.1039/b507237a.
- [160] X. Liu et al., "Biofunctionalized gold nanoparticles for colorimetric sensing of botulinum neurotoxin A light chain," *Anal Chem*, vol. 86, no. 5, pp. 2345-52, Mar 4 2014, doi: 10.1021/ac402626g.
- [161] V. S. Godakhindi et al., "Tuning the Gold Nanoparticle Colorimetric Assay by Nanoparticle Size, Concentration, and Size Combinations for Oligonucleotide Detection," *ACS Sens*, vol. 2, no. 11, pp. 1627-1636, Nov 22 2017, doi: 10.1021/acssensors.7b00482.
- [162] G. Cao, F. Xu, S. Wang, K. Xu, X. Hou, and P. Wu, "Gold Nanoparticle-Based Colorimetric Assay for Selenium Detection via Hydride Generation," (in eng), *Anal Chem*, vol. 89, no. 8, pp. 4695-4700, Apr 18 2017, doi: 10.1021/acs.analchem.7b00337.
- [163] C. D. Medley, J. E. Smith, Z. Tang, Y. Wu, S. Bamrungsap, and W. Tan, "Gold nanoparticle-based colorimetric assay for the direct detection of cancerous cells," (in eng), *Anal Chem*, vol. 80, no. 4, pp. 1067-72, Feb 15 2008, doi: 10.1021/ac702037y.
- [164] P. Wu, K. Hwang, T. Lan, and Y. Lu, "A DNzyme-gold nanoparticle probe for uranyl ion in living cells," (in English), *Journal of the American Chemical Society*, vol. 135, no. 14, pp. 5254-7, Apr 10 2013, doi: 10.1021/ja400150v.
- [165] Y. Guo, S. Li, J. Liu, G. Yang, Z. Sun, and J. Wan, "Double functional aptamer switch probes based on gold nanorods for intracellular ATP detection and targeted drugs transportation," (in English), *Sensors and Actuators B: Chemical*, vol. 235, pp. 655-662, Nov 1 2016, doi: 10.1016/j.snb.2016.05.131.
- [166] R. Elghanian, J. J. Storhoff, R. C. Mucic, R. L. Letsinger, and C. A. Mirkin, "Selective colorimetric detection of polynucleotides based on the distance-dependent optical properties of gold nanoparticles," (in eng), *Science (New York, N.Y.)*, vol. 277, no. 5329, pp. 1078-81, Aug 22 1997, doi: 10.1126/science.277.5329.1078.
- [167] J. J. Storhoff, R. Elghanian, R. C. Mucic, C. A. Mirkin, and R. L. Letsinger, "One-Pot Colorimetric Differentiation of Polynucleotides with Single Base Imperfections Using Gold Nanoparticle Probes," (in English), *Journal of the American Chemical Society*, vol. 120, no. 9, pp. 1959-1964, Mar 11 1998, doi: 10.1021/ja972332i.
- [168] X. Xu, M. S. Han, and C. A. Mirkin, "A gold-nanoparticle-based real-time colorimetric screening method for endonuclease activity and inhibition," (in eng), *Angewandte Chemie (International ed. in English)*, vol. 46, no. 19, pp. 3468-70, 2007, doi: 10.1002/anie.200605249.
- [169] N. Uehara, "Colorimetric assay of homocysteine using gold nanoparticles conjugated with thermoresponsive copolymers," (in English), *Anal Methods-Uk*, vol. 8, no. 39, pp. 7185-7192, 2016, doi: 10.1039/c6ay02002j.
- [170] R. Olafsson, D. R. Bauer, L. G. Montilla, and R. S. Witte, "Real-time, contrast enhanced photoacoustic imaging of cancer in a mouse window chamber," (in English), *Opt Express*, vol. 18, no. 18, pp. 18625-32, Aug 30 2010, doi: 10.1364/OE.18.018625.
- [171] D. Pan, B. Kim, L. V. Wang, and G. M. Lanza, "A brief account of nanoparticle contrast agents for photoacoustic imaging," (in English), *Wiley Interdiscip Rev Nanomed Nanobiotechnol*, vol. 5, no. 6, pp. 517-43, Nov-Dec 2013, doi: 10.1002/wnan.1231.
- [172] D. Pan et al., "Molecular photoacoustic imaging of angiogenesis with integrin-targeted gold nanobeacons," (in English), *Faseb J*, vol. 25, no. 3, pp. 875-82, Mar 2011, doi: 10.1096/fj.10-171728.
- [173] S. Mallidi, G. P. Luke, and S. Emelianov, "Photoacoustic imaging in cancer detection, diagnosis, and treatment guidance," (in English), *Trends Biotechnol*, vol. 29, no. 5, pp. 213-21, May 2011, doi: 10.1016/j.tibtech.2011.01.006.
- [174] J. U. Menon, P. Jadeja, P. Tambe, K. Vu, B. Yuan, and K. T. Nguyen, "Nanomaterials for photo-based diagnostic and therapeutic applications," (in English), *Theranostics*, vol. 3, no. 3, pp. 152-66, 2013, doi: 10.7150/thno.5327.

[175] P. Huang et al., "Biodegradable gold nanovesicles with an ultrastrong plasmonic coupling effect for photoacoustic imaging and photothermal therapy," (in English), *Angewandte Chemie (International ed. in English)*, vol. 52, no. 52, pp. 13958-13964, Dec 23 2013, doi: 10.1002/anie.201308986.

[176] J. J. Richardson and F. Caruso, "Nanomedicine toward 2040," (in eng), *Nano letters*, vol. 20, no. 3, pp. 1481-1482, Mar 11 2020, doi: 10.1021/acs.nanolett.0c00620.

[177] M. Faria et al., "Minimum information reporting in bio-nano experimental literature," (in eng), *Nat Nanotechnol*, vol. 13, no. 9, pp. 777-785, Sep 2018, doi: 10.1038/s41565-018-0246-4.

[178] F. Caputo, J. Clogston, L. Calzolari, M. Rosslein, and A. Prina-Mello, "Measuring particle size distribution of nanoparticle enabled medicinal products, the joint view of EUNCL and NCI-NCL. A step by step approach combining orthogonal measurements with increasing complexity," (in eng), *Journal of controlled release: official journal of the Controlled Release Society*, vol. 299, pp. 31-43, Apr 10 2019, doi: 10.1016/j.jconrel.2019.02.030.

[179] S. Gioria et al., "Are existing standard methods suitable for the evaluation of nanomedicines: some case studies," (in eng), *Nanomedicine-Uk*, vol. 13, no. 5, pp. 539-554, Mar 1 2018, doi: 10.2217/nmm-2017-0338.

[180] J. J. Richardson and F. Caruso, "Nanomedicine toward 2040," (in eng), *Nano Lett*, Mar 2 2020, doi: 10.1021/acs.nanolett.0c00620.