

Safe Bloodstream Concentration Limits for Microscale Nanorobots

Robert A. Freitas Jr.*

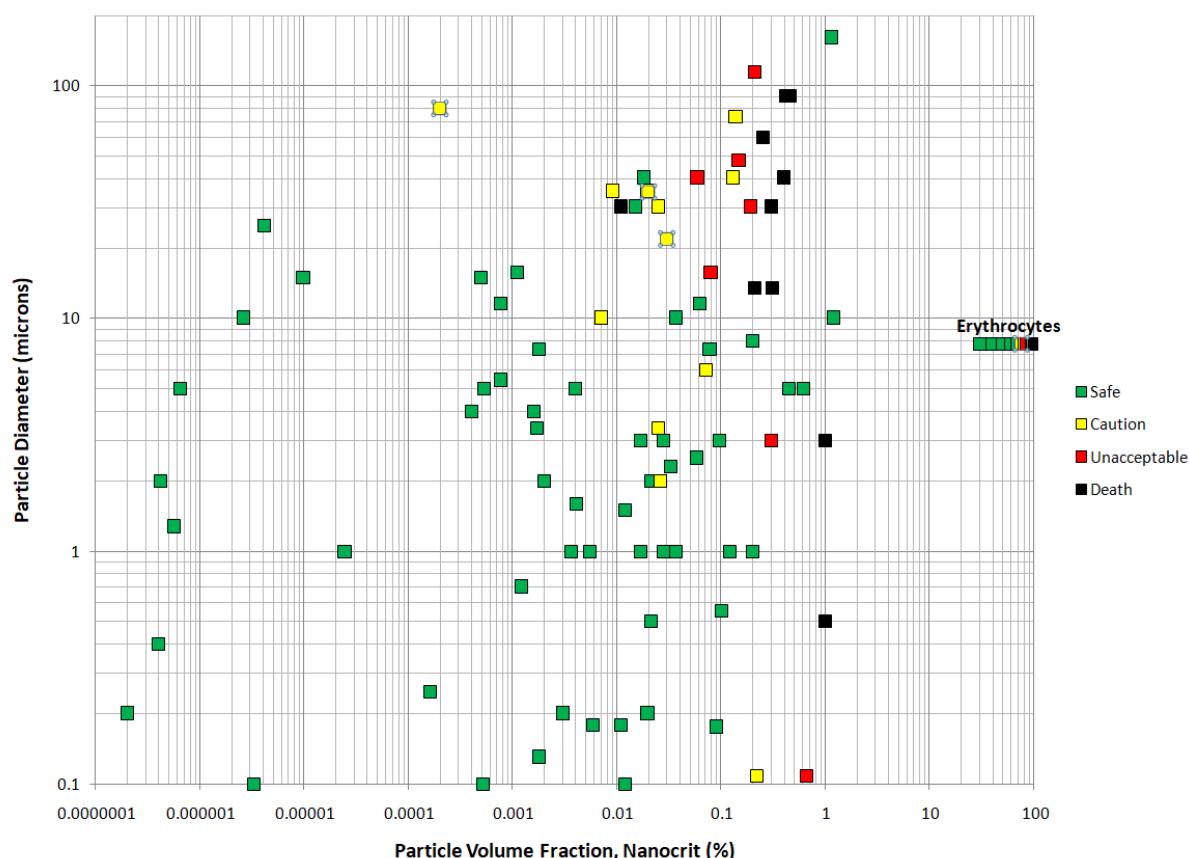
*Institute for Molecular Manufacturing**380 Hamilton Avenue, Suite 112, Palo Alto, CA 94301* Submitted:

February 20, 2025

Accepted: March 10, 2025

Published: March 12, 2025

Graphical Abstract



Abstract

The greatest power of nanomedicine will emerge as we learn to design and construct complete artificial nanorobots using nanometer-scale parts and onboard subsystems such as diamondoid bearings and gears, nanomotors and molecular pumps, nanomanipulators, nanosensors, nanobatteries, and nanocomputers. However, according to initial human intravenous tests of future microscale diamondoid nanorobots in the 0.5-3 μm size range should not exceed a maximum bloodstream volumetric concentration of $\sim 0.001\%$.

Keywords

Nanorobot, bloodstream concentration, safety, nanoparticle, microparticle, diamondoids, humans

Purpose, Rationale, and Limitations

The emergence of atomically precise manufacturing and nano-factories¹ will eventually make possible the design and manufacture of molecular machine systems, including sophisticated micron-scale medical nanorobots,² possi-

bly constructed of atomically precise diamondoids or similar crystalline ceramic materials³ for a wide variety of powerful applications.⁴

Before such robots could be considered for clinical practice, their safety when injected into the bloodstream of animal or human subjects would have to be established, leaving aside the

further analysis of the likely mechanical and kinematic activities of such active-dimensioned devices⁵ and considering them simply as passive rigid microparticles in our initial evaluation.

Introduction

Many experimental studies in animal and human subjects over the last 70 years have directly studied, or have reported as indirect outcomes of related studies, the pathological effects of intravenous injection of microparticles of various sizes and compositions. Definitive safe ranges cannot yet be specified. Still, a maximum safe concentration limit for well-engineered future nanorobots derived from reported experimental results using only the coarsely-shaped passive microparticles surveyed here should provide at least a conservative upper limit. This is because sophisticated future nanorobots will be specifically designed to avoid basic biocompatibility problems⁶ such as accumulation in capillary beds, whereas the passive microparticles in existing studies were not. Additional caution is

warranted because the historical literature's descriptions of outcomes and pathologies have been relatively crude and qualitative.

Many clinical effects have been documented in patients who have received therapeutic injections containing particulate matter contamination,⁷ including infusion-related phlebitis,⁸ pulmonary emboli,⁹ pulmonary granulomas,¹⁰ foreign body reactions from glass particle contaminants,¹¹ immune system dysfunction,¹² pulmonary dysfunction,¹³ adult respiratory distress syndrome,¹⁴ systemic inflammatory response syndrome (SIRS),¹⁵ infarction,¹⁶ and even death.¹⁷ Hard particles $>5\ \mu\text{m}$ in size are more likely to produce mechanical obstruction and vascular emboli.¹⁸ A previously published literature survey¹⁹ found almost exclusive coverage of cell responses to diamondoid particulates but almost no tissue, organ, or whole-body data. Literature data on the bloodstream safety of nontoxic metabolically inert particles are summarized in Figure 1 and Table 1.

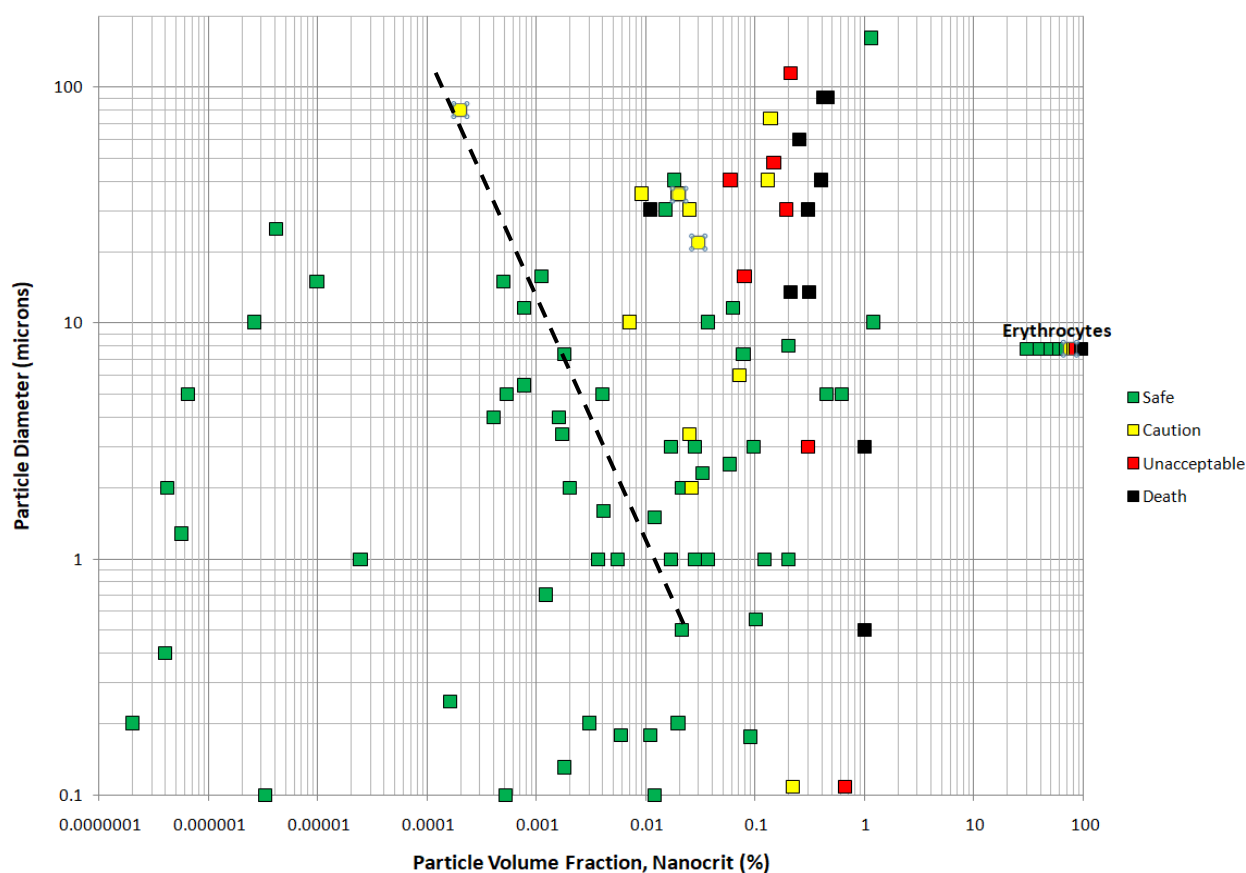


Figure 1. Microparticle safety as a function of diameter and bloodstream concentration

Table 1. Microparticle bloodstream bio-tolerance in animals and humans

Particle Size and Type	Tester	Max. Concentration (Nct)		Reported Consequences
0.2/1/4 μm polystyrene micro-spheres ²⁰	rat	$2 \times 10^{-7} \% / 2.5 \times 10^{-5} \% / 0.0016\%$		no reported discomfort
1.27/15.8 μm polystyrene micro-spheres ²¹	rabbit	$5.6 \times 10^{-7} \% / 0.0011\%$		no adverse effects
>2 μm particle contaminants ²²	human	$4.2 \times 10^{-7} \%$		1980 British Pharmacopoeia: safe
>5 μm particle contaminants ²³	human	$6.5 \times 10^{-7} \%$		1980 British Pharmacopoeia: safe
>10 μm particle contaminants ²⁴	human	$2.6 \times 10^{-6} \%$		1990 British Pharmacopoeia: safe
>25 μm particle contaminants ²⁵	human	$4.1 \times 10^{-6} \%$		1990 British Pharmacopoeia: safe
~15 μm $^{99\text{m}}\text{Tc}$ albumin micro-spheres ²⁶	human	$1 \times 10^{-5} \%$		no toxicity or antibody formation
0.1/0.25 μm gold nanoparticles ²⁷	rat	$3.3 \times 10^{-6} \% / 0.00016\%$		no reported discomfort
0.11 μm silver nanoparticles ²⁸	rat	$1.5 \times 10^{-5} \%$		no adverse effects
10 x 80 μm cotton filter paper ²⁹	rabbit	0.0002%		some granuloma formation
0.4/4 μm polystyrene latex ³⁰	rat	$4 \times 10^{-7} \% / 0.0004\%$		well tolerated
~15 μm $^{99\text{m}}\text{Tc}$ human albumin micro-spheres ³¹	rat	0.0005%		no reported discomfort
0.1 μm nanodiamonds ³²	rat	0.00052%		no reported discomfort
5 μm DEAE-cellulose fibers ³³	rabbit	0.00053%		no adverse effects
11.6 μm polystyrene microspheres ³⁴	dog	0.00076%		no reported discomfort
5.4 μm polystyrene microspheres ³⁵	dog	0.00077%		no reported discomfort
0.7/1/1.5/2.5/3/5 μm spherical silica beads ³⁶	mouse	0.0012% / 0.0036% / 0.012% / 0.058% / 0.097% / 0.45%		no reported discomfort
3.4 μm polystyrene microspheres ³⁷	dog	0.0017%		no reported discomfort
7.4 μm polystyrene microspheres ³⁸	dog	0.0018%		no reported discomfort
0.13 μm gold-silica nanoshells ³⁹	mouse	0.0018%		all mice remained healthy
2/5 μm bubbles ⁴⁰	human	0.002% / 0.004%		well-tolerated
~0.2 μm polymersomes ⁴¹	mouse	0.003%		maximum tolerated dose
1.6 μm hemispherical silica beads ⁴²	mouse	0.0041%		no reported discomfort
1.6 μm discoidal silica beads ⁴³	mouse	0.0041%		no reported discomfort
1 μm cylindrical silica beads ⁴⁴	mouse	0.0055%		no reported discomfort
0.05-0.18 μm nanodiamonds ⁴⁵	rat	0.0059%		no adverse effects
~35 μm macroaggregated albumin ⁴⁶	monkey (brain)	0.006%		no behavioral effects
10 μm polystyrene latex ⁴⁷	rat	0.007%		focal myocardial degeneration onset
~35 μm macroaggregated albumin ⁴⁸	monkey (brain)	0.009%		onset of focal necrosis
0.05-0.18 μm nanodiamonds ⁴⁹	monkey	0.011%		no adverse effects
30 μm DEAE-cellulose fibers ⁵⁰	rabbit	0.011%		~LD ₅₀ (acute toxic response)

0.1/1/5/10 µm spheres & 0.1x1x3 µm disks, of polystyrene ⁵¹	mouse	0.012% / 0.12% / 0.61% / 1.2% / 0.033%		no reported discomfort
30 µm albumin microspheres ⁵²	dog	0.015%		maximum nontoxic dose
	dog	0.025%		evidence of toxicity
	dog	0.19%		pulmonary-artery pressure+300%
	dog	~0.3%		lethal
1-3 µm alumina ⁵³	rabbit	0.017%		no evidence of disease
40/160 µm DEAE-cellulose microspheres ⁵⁴	rabbit	0.018% / 1.13%		no adverse effects
~35 µm macroaggregated albumin ⁵⁵	dog	0.02%		tachypnea
~0.2 µm PEG nanoparticles ⁵⁶	mouse	0.02%		no reported discomfort
0.5-2 µm SWCNTs ⁵⁷	mouse	0.021%		no reported discomfort
3.4 µm polystyrene microspheres ⁵⁸	beagle	0.025%		systemic hypotension & myocardial depression
2/6 µm cationic microbubbles ⁵⁹	rat	0.026% / 0.071%		minimal hemorrhage
1-3 µm diamond dust ⁶⁰	rabbit	0.028%		no evidence of disease
22 µm albumin spheres ⁶¹	dog	0.03%		pulmonary-artery pressure +42%
1-10 µm MWCNTs ⁶²	mouse	0.037%		no reported discomfort
40 µm starch ⁶³	human	0.06%		50% nausea and vomiting
7.4/11.6 µm polystyrene microspheres ⁶⁴	beagle	0.079% / 0.062%		no significant hemodynamic effect
15.8/48/115 µm polystyrene/latex ⁶⁵	dog	0.081% / 0.15% / 0.21%		pulmonary-artery pressure+100%
~0.175 µm PEG-PE liposomes ⁶⁶	mouse	0.09%		no reported discomfort
~0.1 µm colloidal carbon ⁶⁷	rabbit	0.1%		no reported discomfort
0.557 µm polystyrene latex ⁶⁸	rabbit	0.1%		no reported discomfort
40 µm cotton fibers ⁶⁹	rat	0.13%		some granuloma formation
74 µm cotton filter paper ⁷⁰	dog	0.14%		numerous pulmonary granulomas
1/8 µm carbon ⁷¹	rat	0.2% / 0.2%		no reported discomfort
0.11 µm Mg ₂ Al(OH) ₆ Cl “LDH” ⁷²	rat	0.22%		inflammation & some necrosis
0.11 µm Mg ₂ Al(OH) ₆ Cl “LDH” ⁷³	rat	0.66%		lesions, edema & necrosis
3 µm latex ⁷⁴	beagle	0.3%		dyspnea
40 µm polystyrene latex ⁷⁵	rat	0.4%		~LD ₅₀
13.5/90.7 µm latex ⁷⁶	mouse	0.21% / 0.46%		24-hr LD ₅₀
60-420 µm glass beads ⁷⁷	dog	0.25%		~LD ₅₀
13.5/90.7 µm latex ⁷⁸	rat	0.31% / 0.43%		24-hr LD ₅₀
3 µm latex ⁷⁹	rat	~1%		est. LD ₅₀
0.5 µm <i>S. aureus</i> bacteria ⁸⁰	mouse	~1%		~LD ₅₀
7-8 µm erythrocytes	human	28%-71%		normal range
7-8 µm erythrocytes	human	>71%		polycythemia
7-8 µm erythrocytes	human	85%		likely stroke/heart failure
7-8 µm erythrocytes	human	94%		death from stroke/heart failure

See color codes in Figure 1.

Discussion

Historical data suggests that initial human intravenous tests of microscale diamondoid nanorobots in the 0.5-3 μm size range probably should not exceed a maximum target concentration of $N_{\text{ct}} \sim 0.001\%$ (e.g., ~ 13 billion 2- μm spherical nanorobots, of total robot volume $V_{\text{robots}} \sim 54 \text{ mm}^3$, in an adult human male with $V_{\text{blood}} \sim 5.4$ liters of circulating whole blood), where $V_{\text{robots}} / V_{\text{blood}} = N_{\text{ct}}$ (“nanocrit”) is the percentage volume concentration of nanorobots in the blood. Up to $N_{\text{ct}} \sim 0.005\%$ might still be considered acceptably conservative, given the dashed line in Figure 1 representing the empirical “safe” boundary of $D_{\text{sph}} (\mu\text{m}) \times N_{\text{ct}} (\%) \leq 0.01$.

The human LD_{50} for freely-circulating micro-particles of any kind has not been reported in the literature. Microsphere toxicity in animals due to vascular occlusion is a function of the total volume of microspheres injected.⁸¹ For rats appears to follow a power law of the form $N_{\text{LD50, rat}} = M_{\text{body}} K / D_{\text{sph}}^b$, where $N_{\text{LD50, rat}}$ is the number of spheres intravenously injected to achieve LD_{50} in the rat, M_{body} is rat body weight in kg, D_{sph} = microsphere diameter in microns, and the experimentally-determined constants are $K \sim 8.6 \times 10^{11} \text{ micron}^3/\text{kg}$ and $b \sim 3.15$; with a similar relation found for dogs.⁸² Simply extrapolating to a human body weight of $M_{\text{body}} = 70 \text{ kg}$, the LD_{50} whole-body LD_{50} dose of $D_{\text{sph}} = 1 \text{ micron}$ latex microspheres would be $N_{\text{LD50, human}} \sim 60$ trillion spheres, or $N_{\text{ctLD50, human}} \sim (\pi/6) D_{\text{sph}}^3 N_{\text{LD50, human}} / V_{\text{blood}} \sim 0.6\%$. This

might be considered a pessimistic estimate because the above formula for N_{LD50} is derived mainly from studies of (1) experimental particles that are >10 microns in diameter, whereas proposed bloodborne microscale nanorobots are typically 0.5-3 μm , (2) particles having varying dimensions and atomically rough surfaces, whereas proposed atomically precise nanorobots can have identical radii and atomically smooth and engineered-biocompatible surfaces, and (3) particles that were deployed in animal models (e.g., rats and rabbits) that have smaller average red blood cell diameters than humans. On the other hand, the estimate might be considered optimistic for continuously circulating nanorobots because the aforementioned experimental particles are rapidly cleared from the bloodstream via the liver, spleen, kidney, and lungs, reducing their actual experimental dwell time in the circulation.

It is worth noting that natural erythrocytes, if regarded as superbly engineered biological microscale nanomachines, safely achieve the equivalent of $N_{\text{ct}} \sim 13\%$ in 8- μm capillaries, $N_{\text{ct}} \sim 25\%$ in 30- μm pre-capillary channels, $N_{\text{ct}} \sim 36\%$ in 100- μm arterioles and venules, and $N_{\text{ct}} = 46\%$ (range 43%-58%) in the largest vessels of the normal human vasculature.⁸³ This suggests the possibility of at least a 100-1000 fold improvement in the maximum tolerable bloodstream concentration of microscale nanorobots if they are equipped with comparably engineered biocompatible exteriors, as compared to the raw-surfaced and coarsely-shaped passive microparticles surveyed here.

Conclusions

A survey of experimental studies on the pathological effects of intravenous injection of microparticles of various sizes and compositions, conducted over the last 70 years in both animal and human subjects, suggests that initial human intravenous tests of microscale diamondoid nanorobots in the 0.5-3 μm size range should not exceed a maximum bloodstream volumetric concentration of approximately 0.001%.

Funding: The research in this paper was supported by the Institute for Molecular Manufacturing.

Conflicts of Interest: The author declares no conflict of interest. For a signed statement, contact the Editorial Office.

Quote this article as Robert A. Freitas Jr., Safe Bloodstream Concentration Limits for Microscale Nanorobots, *Precis. Nanomed.* 2025, 8(1):1454-1463, <https://doi.org/10.33218/001c.132323>.

COPYRIGHT NOTICE ©The Author(s) 2024. This article is distributed under the terms of the [Creative Commons Attribution 4.0 International License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

1. Drexler KE. Molecular engineering: An approach to the development of general capabilities for molecular manipulation. *Proc Natl Acad Sci U S A*. 1981 Sep;78(9):5275-8; <https://www.pnas.org/doi/pdf/10.1073/pnas.78.9.5275>. Drexler KE. *Nanosystems: Molecular Machinery, Manufacturing, and Computation*, John Wiley & Sons, New York, 1992; http://e-drexler.com/d/09/00/Drexler_MIT_dissertation.pdf. Freitas RA Jr., Merkle RC. *Kinematic Self-Replicating Machines*. Landes Bioscience, Georgetown, TX, 2004; <http://www.MolecularAssembler.com/KSRM.htm>
2. Freitas RA Jr. *Nanomedicine, Volume I: Basic Capabilities*. Landes Bioscience, Georgetown, TX, 1999; <http://www.nanomedicine.com/NMI.htm>.
3. Freitas RA Jr. *Nanomedicine, Volume IIA: Biocompatibility*, Landes Bioscience, Georgetown, TX, 2003; <http://www.nanomedicine.com/NMIIA.htm>.
4. Freitas RA Jr. Chapter 23. *Comprehensive Nanorobotic Control of Human Morbidity and Aging*. In: Fahy GM, West MD, Coles LS, Harris SB, eds, *The Future of Aging: Pathways to Human Life Extension*, Springer, New York, 2010; <http://www.nanomedicine.com/Papers/Aging.pdf>.
5. Freitas RA Jr. Exploratory design in medical nanotechnology: a mechanical artificial red cell. *Artif Cells Blood Substit Immobil Biotechnol*. 1998 Jul;26(4):411-30; <https://www.tandfonline.com/doi/pdf/10.3109/10731199809117682>. Freitas RA Jr. Microbi-vores: Artificial mechanical phagocytes using digest and discharge protocol. *J Evol Technol* 2005;14:1-52; <http://www.jetpress.org/volume14/freitas.html>. Freitas RA Jr. The ideal gene delivery vector: Chromalloyocytes, cell repair nanorobots for chromosome replacement therapy. *J Evol Technol* 2007;16:1-97; <http://jetpress.org/v16/freitas.pdf>.
6. Freitas RA Jr., 2003.
7. Barber TA. Chapter 1. Introduction and Overview. In: *Control of Particulate Matter: Contamination in Healthcare Manufacturing*; Interpharm Press: Buffalo Grove, IL, 2000, pp 1-26.
8. Pesko LJ. Physiological Consequences of Injected Particulates. In: *Liquid and Surface Borne Particle Measurement Handbook*; Marcel Dekker, Inc., New York, 1996; pp 661-686. Falchuk KH, Peterson L, McNeil BJ. Microparticulate-induced phlebitis. Its prevention by in-line filtration. *N Engl J Med*. 1985 Jan 10;312(2):78-82; <https://www.ncbi.nlm.nih.gov/pubmed/3880597>. Tran T, Kupiec TK, Trissel LA. Quality-Control Analytical Methods: Particulate Matter in Injections: What is it and What are the Concerns? *Int J Pharm Compd*. 2006 May-Jun;10(3):202-4; <https://www.ncbi.nlm.nih.gov/pubmed/23974233>. Gurgvich I. Letter to the Editor. *New Engl. J. Med*. 1985;312(22):1453. Kirkpatrick CJ, Rangoonwala R. Originator antibiotic or generic in clinical practice. *J. Appl. Ther. Res*. 2009;7:52-57.
9. Hill SE, Heldman LS, Goo ED, Whippo PE, Perkinson JC. Fatal microvascular pulmonary emboli from precipitation of a total nutrient admixture solution. *JPEN J Parenter Enteral Nutr*. 1996 Jan-Feb;20(1):81-7; <https://www.ncbi.nlm.nih.gov/pubmed/8788269>. Schaefer SC, Bison PA, Rangoonwala R, Kirkpatrick CJ, Lehr HA. 0.2 μ m in-line filters prevent capillary obstruction by particulate contamination of generic antibiotic preparations in postischemic muscle. *Chemother. J*. 2008;17:172-178. Illum L, Davis SS, Wilson CG, Thomas NW, Frier M, Hardy JG. Blood Clearance and Organ Deposition of Intravenously Administered Colloidal Particles – the Effects of Particle-Size, Nature and Shape. *Int. J. Pharm*. 1982;12(2-3):135-146; <https://www.sciencedirect.com/science/article/pii/0378517382901132>.
10. Malizia AA Jr, Reiman HM, Myers RP, Sande JR, Barham SS, Benson RC Jr, Dewanjee MK, Utz WJ. Migration and granulomatous reaction after periurethral injection of polytetrafluoroethylene (Teflon). *JAMA*. 1984 Jun 22-29;251(24):3277-81; <https://www.ncbi.nlm.nih.gov/pubmed/6374180>. Puntis JW, Wilkins KM, Ball PA, Rushton DI, Booth IW. Hazards of parenteral treatment: do particles count? *Arch Dis Child*. 1992 Dec;67(12):1475-7; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1793986/>. Pesko, 1996. Tran *et al.*, 2006.
11. Sabon RL Jr, Cheng Er, Stommel KA, Hennen CR. Glass particle contamination: influence of aspiration methods and ampoule types. *Anesthesiology* 1989 May; 70(5):859-62; <https://www.ncbi.nlm.nih.gov/pubmed/2719321>. Lye ST, Hwang NC. Glass particle contamination

- tion: Is it here to stay? *Anesthesia* 2003;58(1):93-94; <https://onlinelibrary.wiley.com/doi/full/10.1046/j.1365-2044.2003.296812.x>. Zabir AF, Choy CY, Bushdan R. Glass particle contamination of parenteral preparations of intravenous drugs in anaesthetic practice. *Southern African Journal of Anaesthesia and Analgesia* 2008;14(3):17-19; <https://pdfs.semanticscholar.org/f336/c0e6903717088cbff85fc5c9b7c12448f3b.pdf>.
12. Pesko, 1996. Jack T, Brent BE, Boehne M, Müller M, Sewald K, Braun A, Wessel A, Sasse M. Analysis of particulate contaminations of infusion solutions in a pediatric intensive care unit. *Intensive Care Med.* 2010 Apr;36(4):707-11; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2837187/>.
 13. ¹ Kirkpatrick CJ, Rangoonwala R. Originator antibiotic or generic in clinical practice. *J. Appl. Ther. Res.* 2009;7:52-57. Schaefer SC, Bison PA, Rangoonwala R, Kirkpatrick CJ, Lehr HA. 0.2 µm in-line filters prevent capillary obstruction by particulate contamination of generic antibiotic preparations in postischemic muscle. *Chemother. J.* 2008;17:172-178.
 14. Walpot H, Franke RP, Burchard WG, Agternkamp C, Müller FG, Mittermayer C, Kalff G. [Particulate contamination of infusion solutions and drug additives within the scope of long-term intensive therapy. 1. Energy dispersion electron images in the scanning electron microscope-REM/EDX]. *Anaesthesist.* 1989 Oct;38(10):544-8 [in German]; <https://www.ncbi.nlm.nih.gov/pubmed/2511778>.
 15. Brauser D. Inline filtration may reduce SIRS mortality in the pediatric ICU. 14 Jan 2010; <http://www.medscape.com/viewarticle/715108>
 16. Winding O, Grønvald J, Faarup P, Hegedüs V. Sequelae of intrinsic foreign-body contamination during selective renal angiography in rabbits. *Radiology.* 1980 Feb;134(2):321-6; <https://www.ncbi.nlm.nih.gov/pubmed/7352209>. Schaefer *et al.*, 2008.
 17. Hill SE, Heldman LS, Goo ED, Whippo PE, Perkinson JC. Fatal microvascular pulmonary emboli from precipitation of a total nutrient admixture solution. *JPEN J Parenter Enteral Nutr.* 1996 Jan-Feb;20(1):81-7; <https://www.ncbi.nlm.nih.gov/pubmed/8788269>. Shay DK, Fann LM, Jarvis WR. Respiratory distress and sudden death associated with receipt of a peripheral parenteral nutrition admixture. *Infect Control Hosp Epidemiol.* 1997 Dec;18(12):814-7; <https://www.ncbi.nlm.nih.gov/pubmed/9442405>. Bradley JS, Wassel RT, Lee L, Nambiar S. Intravenous ceftriaxone and calcium in the neonate: assessing the risk for cardiopulmonary adverse events. *Pediatrics.* 2009 Apr;123(4):e609-13; <https://www.ncbi.nlm.nih.gov/pubmed/19289450>. Cant AJ, Lenney W, Kirkham N. Plastic material from a syringe causing fatal bowel necrosis in a neonate. *Br Med J (Clin Res Ed).* 1988 Apr 2;296(6627):968-9; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2545438/>.
 18. Langille SE. Particulate matter in injectable drug products. *PDA J Pharm Sci Technol.* 2013 May-Jun;67(3):186-200; <https://citeseerx.ist.psu.edu/document?repid=rep1&type=pdf&doi=dfb66f05f244a551995fd8391b69a3a53111e67>.
 19. Freitas RA Jr. Nanomedicine, Volume IIA: Biocompatibility, Landes Bioscience, Georgetown, TX, 2003; Section 15.3.1.4, "Biocompatibility of Diamond Particles"; <http://www.nanomedicine.com/NMIIA/15.3.1.4.htm>.
 20. Yokel RA, Sabo JP, Simmons GH, DeLuca PP. Acute toxicity of latex microspheres. *Toxicol Lett.* 1981 Oct;9(2):165-70; <https://www.ncbi.nlm.nih.gov/pubmed/7302990>.
 21. Allen DR, Ferens JM, Cheney FW, Nelp WB. Critical evaluation of acute cardiopulmonary toxicity of microspheres. *J. Nucl. Med.* 1978 Nov;19:1204-1208; <https://jnm.snmjournals.org/content/jnumed/19/11/1204.full-text.pdf>.
 22. Barbee JH, Cokelet GR. The Fahraeus effect. *Microvasc Res.* 1971 Jan;3(1):6-16; <https://pubmed.ncbi.nlm.nih.gov/5092929/>. Barbee JH, Cokelet GR. Prediction of blood flow in tubes with diameters as small as 29 microns. *Microvasc Res.* 1971 Jan;3(1):17-21; <https://www.sciencedirect.com/science/article/abs/pii/0026286271900033>.
 23. Gao QP, Hansen B, Seljelid R. Influence of Size, Dosage, and Surface Structure on Clearance and Tissue Distribution of Intravenous Microspheres, *Drug Delivery* 1997; 4(2):93-99; <https://www.tandfonline.com/doi/pdf/10.3109/10717549709051879>.

24. Ilium L, Davis SS, Wilson CG, Thomas NW, Frier M, Hardy JG. Blood clearance and organ deposition of intravenously administered colloidal particles. The effects of particle size, nature and shape. *Intl. J Pharmaceutics* 1982; 12(2-3):135-146; [https://doi.org/10.1016/0378-5173\(82\)90113-2](https://doi.org/10.1016/0378-5173(82)90113-2), <https://www.sciencedirect.com/science/article/pii/0378517382901132>
25. The British Pharmacopoeia 1980, The Pharmaceutical Press, London, 1980. Cited in: Madsen H, Winding O. Release of foreign bodies (particles) by clinical use of intravenous infusion sets. *Bio-materials*. 1996 Apr;17(7):663-6; <https://www.ncbi.nlm.nih.gov/pubmed/8672627>, <https://www.sciencedirect.com/science/article/pii/0142961296867354>.
26. British Pharmacopoeia 1980.
27. The United States Pharmacopoeia 1990. Cited in: Madsen and Winding, 1996.
28. British Pharmacopoeia 1990
29. Burdine JA, Ryder LA, Sonnemaker RE, DePuey G, Calderson M. ^{99m}Tc-human albumin microspheres (HAM) for lung imaging. *J Nucl Med*. 1971 Mar;12(3):127-30; <http://jnm.snmjournals.org/content/12/3/127.long>.
30. De Jong WH, Hagens WI, Krystek P, Burger MC, Sips AJ, Geertsma RE. Particle size-dependent organ distribution of gold nanoparticles after intravenous administration. *Biomaterials*. 2008 Apr;29(12):1912-9; <https://www.ncbi.nlm.nih.gov/pubmed/18242692>.
31. Lankveld DP, Oomen AG, Krystek P, Neigh A, Troost-de Jong A, Noorlander CW, Van Eijkeren JC, Geertsma RE, De Jong WH. The kinetics of the tissue distribution of silver nanoparticles of different sizes. *Biomaterials*. 2010 Nov;31(32):8350-61; https://www.academia.edu/download/51611430/The_kinetics_of_the_tissue_distribution_20170202-5327-16p9y1r.pdf.
32. Wartman WB, Hudson B, Jennings RB. Experimental arterial disease. II. The reaction of the pulmonary artery to emboli of filter paper fibers. *Circulation*. 1951 Nov;4(5):756-63; <http://circ.ahajournals.org/content/circulationaha/4/5/756.full.pdf>.
33. Gesler RM, Garvin PJ, Klamer B, Robinson RU, Thompson CR, Gibson WR, Wheeler FC, Carlson RG. The biologic effects of polystyrene latex particles administered intravenously to rats – a collaborative study. *Bull Parenter Drug Assoc*. 1973 May-Jun;27(3):101-13; <https://www.ncbi.nlm.nih.gov/pubmed/4709158>.
34. Burdine *et al*, 1971.
35. Tsai LW, Lin YC, Perevedentseva E, Lugovtsov A, Priezzhev A, Cheng CL. Nanodiamonds for Medical Applications: Interaction with Blood in Vitro and in Vivo. *Int J Mol Sci*. 2016 Jul 12;17(7); <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4964486/>.
36. Ilium *et al.*, 1982.
37. Kanke M, Simmons GH, Weiss DL, Bivins BA, DeLuca PP. Clearance of ¹⁴¹Ce-labeled microspheres from blood and distribution in specific organs following intravenous and intraarterial administration in beagle dogs. *J Pharm Sci*. 1980 Jul;69(7):755-62; <https://onlinelibrary.wiley.com/doi/full/10.1002/jps.2600690703>.
38. Kanke *et al.*, 1980.
39. Decuzzi P, Godin B, Tanaka T, Lee SY, Chiappini C, Liu X, Ferrari M. Size and shape effects in the biodistribution of intravascularly injected particles. *J Control Release*. 2010 Feb 15;141(3):320-7; <http://chiappinilab.com/wp-content/uploads/2016/08/J.-Control.-Rel.-2010-Decuzzi.pdf>.
40. Kanke *et al.*, 1980
41. Kanke *et al.*, 1980
42. O'Neal DP, Hirsch LR, Halas NJ, Payne JD, West JL. Photo-thermal tumor ablation in mice using near infrared-absorbing nanoparticles. *Cancer Lett*. 2004 Jun 25;209(2):171-6; <https://citeseerx.ist.psu.edu/document?repid=rep1&type=pdf&doi=463d05701ef81e67042a75a646a8140f36f64abb>.
43. Wells P. Chapter 7. Ultrasonic Contrast Agents. In: Coombs RRH, Robinson DW, eds., *Nanotechnology in Medicine and the Biosciences*, Gordon & Breach Publishers, Netherlands, 1996, pp. 111-121. Barnhart JL, Levene HB, Keen R, Saravis C, Maniquis J, Villapando E. Alunex microsphere shells, the unique feature of a new ultrasound contrast agent. In: Katayama H, Brasch RC, eds., *New Dimensions of Contrast Media*, Excerpta Medica, Amsterdam, 1991. Feinstein

- SB, Cheirif J, Ten Cate FJ, Silverman PR, Heidenreich PA, Dick C, Desir RM, Armstrong WF, Quinones MA, Shah PM. Safety and efficacy of a new transpulmonary ultrasound contrast agent: 44. Ahmed F, Pakunlu RI, Srinivas G, Brannan A, Bates F, Klein ML, Minko T, Discher DE. Shrinkage of a rapidly growing tumor by drug-loaded polymersomes: pH-triggered release through copolymer degradation. *Mol Pharm*. 2006 May-Jun;3(3):340-50; https://discherlab.seas.upenn.edu/wp-content/uploads/2023/06/Psomes_ShrinkTumors-ACS-MoTherapeutics.pdf.
45. Decuzzi *et al.*, 2010.
46. Decuzzi *et al.*, 2010.
47. Decuzzi *et al.*, 2010
48. Moore L, Yang J, Lan TT, Osawa E, Lee DK, Johnson WD, Xi J, Chow EK, Ho D. Biocompatibility Assessment of Detonation Nanodiamond in Non-Human Primates and Rats Using Histological, Hematologic, and Urine Analysis. *ACS Nano*. 2016 Aug 23;10(8):7385-400; <https://www.ncbi.nlm.nih.gov/pubmed/27439019>.
49. Taplin GV, Macdonald NS. Radiochemistry of macroaggregated albumin and newer lung scanning agents. *Semin Nucl Med* 1:132-152, 1971; <https://kundoc.com/pdf-radiochemistry-of-macroaggregated-albumin-and-newer-lung-scanning-agents-.html>.
50. Gessler *et al.*, 1973.
51. Taplin and Macdonald, 1971.
52. Moore *et al.*, 2016.
53. Ilium *et al.*, 1982.
54. Muro S, Garnacho C, Champion JA, Leferovich J, Gajewski C, Schuchman EH, Mitragotri S, Muzykantov VR. Control of endothelial targeting and intracellular delivery of therapeutic enzymes by modulating the size and shape of ICAM-1-targeted carriers. *Mol Ther*. 2008 Aug;16(8):1450-8; <https://pmc.ncbi.nlm.nih.gov/articles/PMC2810502/>
55. Allen DR, Nelp WB, Cheney F, Hartnett DE. Studies of acute cardiopulmonary toxicity of Sn-macroaggregated albumin in the dog. *J Nucl Med*. 1974 Jul;15(7):567-71; <https://jnm.snmjournals.org/content/jnumed/15/7/567.full-text.pdf>.
56. Gardner LU, Cummings DE. The Reaction to Fine and Medium Sized Quartz and Aluminum Oxide Particles. Silicotic Cirrhosis of the Liver. *Am J Pathol*. 1933;9(Suppl):751-764.5; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2062773/pdf>.
57. Ilium *et al.*, 1982.
58. Taplin and Macdonald, 1971.
59. Gratton SE, Pohlhaus PD, Lee J, Guo J, Cho MJ, Desimone JM. Nanofabricated particles for engineered drug therapies: a preliminary biodistribution study of PRINT nanoparticles. *J Control Release*. 2007 Aug 16;121(1-2):10-18; <https://pmc.ncbi.nlm.nih.gov/articles/PMC1994820/pdf>.
60. Singh R, Pantarotto D, Lacerda L, Pastorin G, Klumpp C, Prato M, Bianco A, Kostarelos K. Tissue biodistribution and blood clearance rates of intravenously administered carbon nanotube radiotracers. *Proc Natl Acad Sci U S A*. 2006 Feb 28;103(9):3357-62; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1413890/>.
61. Slack JD, Kanke M, Simmons GH, DeLuca PP. Acute hemodynamic effects and blood pool kinetics of polystyrene microspheres following intravenous administration. *J Pharm Sci*. 1981 Jun;70(6):660-4; <https://pubmed.ncbi.nlm.nih.gov/7252811/>
62. Song KH, Fan AC, Hinkle JJ, Newman J, Borden MA, Harvey BK. Microbubble gas volume: A unifying dose parameter in blood-brain barrier opening by focused ultrasound. *Theranostics*, 2017 Jan 1;7(1):144-152; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5196892/>.
63. Gardner and Cummings, 1933
64. Mishkin FS, Brashear RE. Pulmonary and systemic blood pressure responses to large doses of albumin microspheres. *J. Nucl. Med*. 1971 May;12:251-252; <http://jnm.snmjournals.org/content/12/5/251.full.pdf>.
65. Wei Q, Zhan L, Juanjuan B, Jing W, Jianjun W, Taoli S, Yi'an G, Wangsuo W. Biodistribution of co-exposure to multi-walled carbon nanotubes and nanodiamonds in mice. *Nanoscale Res Lett*. 2012 Aug 23;7(1):473; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3478214/>

66. Ensminger WD, Gyves JW, Stetson P, Walker-Andrews S. Phase I study of hepatic arterial degradable starch microspheres and mitomycin. *Cancer Res.* 1985 Sep;45(9):4464-7; <http://cancerres.aacrjournals.org/content/canres/45/9/4464.full.pdf>. Gyves JW, Ensminger WD, VanHarken D, Niederhuber J, Stetson P, Walker S. Improved regional selectivity of hepatic arterial mitomycin by starch microspheres. *Clin Pharmacol Ther.* 1983 Aug;34(2):259-65; <https://deepblue.lib.umich.edu/bitstream/handle/2027.42/109929/cptclpt1983163.pdf>. Dakhil S, Ensminger W, Cho K, Niederhuber J, Doan K, Wheeler R. Improved regional selectivity of hepatic arterial BCNU with degradable microspheres. *Cancer.* 1982 Aug 15;50(4):631-5; <https://onlinelibrary.wiley.com/doi/pdf/10.1002/1097-0142%2819820815%2950%3A4%3C631%3A%3AAID-CNCR2820500403%3E3.0.CO%3B2-M>.
67. Slack *et al.*, 1981.
68. Allen *et al.*, 1978.
69. Litzinger DC, Buiting AM, van Rooijen N, Huang L. Effect of liposome size on the circulation time and intraorgan distribution of amphipathic poly(ethylene glycol)-containing liposomes. *Biochim Biophys Acta.* 1994 Feb 23;1190(1):99-107; <https://pubmed.ncbi.nlm.nih.gov/8110825/>.
70. Burke JS, Simon GT. Electron microscopy of the spleen. II. Phagocytosis of colloidal carbon. *Am J Pathol.* 1970 Jan;58(1):157-81; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2032827/pdf>.
71. Schoenberg MD, Gilman PA, Mumaw VR, Moore RD. The phagocytosis of uniform polystyrene latex particles (PLP) by the reticulo endothelial system (RES) in the rabbit. *Br J Exp Pathol.* 1961 Oct;42:486-95; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2082491/pdf>.
72. Von Glahn WC, Hall JW. The reaction produced in the pulmonary arteries by emboli of cotton fibers. *Am J Pathol.* 1949 Jul;25(4):575-95; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1942825/pdf>.
73. Gross MA. The Danger of Particulate Matter: In Solutions for Intravenous Use. *Drug Intell.* 1967 Jan 1;1:12; <http://journals.sagepub.com/doi/abs/10.1177/106002806700100103?journalCode=aopa>.
74. Helbing G, Burri C, Mohr W, Neugebauer R, Wolter D. Chapter 38. The Reaction of Tissue to Carbon Particles. In: Winter GD, Leray JL, de Groot K, eds., *Evaluation of Biomaterials*, John Wiley & Sons, New York, 1977, pp. 373-380.
75. Kwak SY, Kriven WM, Wallig MA, Choy JH. Inorganic delivery vector for intravenous injection. *Biomaterials.* 2004 Dec;25(28):5995-6001; <https://pubmed.ncbi.nlm.nih.gov/15183614/>.
76. Kwak *et al.*, 2004.
77. Yokel *et al.*, 1981.
78. Gessler *et al.*, 1973.
79. Davis MA, Taube RA. Pulmonary perfusion imaging: acute toxicity and safety factors as a function of particle size. *J. Nucl. Med.* 1978 Nov;19:1209-1213; <http://jnm.snmjournals.org/content/19/11/1209.short>.
80. Niden AH, Aviado DM Jr. Effects of pulmonary embolism on the pulmonary circulation with special reference to arteriovenous shunts in the lung. *Circ Res.* 1956 Jan;4(1):67-73; <https://pdfs.semanticscholar.org/4d48/277efc356d4eed248395b066fc5056535935.pdf>
81. Davis and Taube, 1978.
82. Yokel *et al.*, 1981.
83. Hasegawa N, Kondo I. Isolation and virulence of a caseinase- and bound coagulase-deficient mutant of *Staphylococcus aureus* BB. *J Infect Dis.* 1984 Apr;149(4):538-43; <https://www.ncbi.nlm.nih.gov/pubmed/6373962>. Bogni C, Segura M, Giraudo J, Giraudo A, Calzolari A, Nagel R. Avirulence and immunogenicity in mice of a bovine mastitis *Staphylococcus aureus* mutant. *Can J Vet Res.* 1998 Oct;62(4):293-8; <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1189497/pdf>.